

ಮಂಗಳೂರು ವಿಶ್ವವಿದ್ಯಾನಿಲಯ
MANGALORE UNIVERSITY



(Accredited by NAAC with 'A' Grade)

ಕ್ರಮಾಂಕ/ No. : MU/ACC/CR 41/2020-21/A2

ಕುಲಸಚಿವರ ಕಛೇರಿ

ಮಂಗಳಗಂಗೋತ್ರಿ - 574 199

Office of the Registrar

Mangalagangothri - 574 199

ದಿನಾಂಕ/Date:15.01.2021

NOTIFICATION

Sub: Revised syllabus of M.Sc. Biosciences programme.

Ref: Academic Council approval vide agenda

No.: ಎ.ಸಿ.ಸಿ.ಶೈ.ಸಾ.ಸ.2:19(2020-21)dtd 23.12.2020.

The Revised syllabus of M.Sc. Biosciences programme which is approved by the Academic Council at its meeting held on 23.12.2020 is hereby notified for implementation with effect from the academic year 2020-21.

Copy of the Syllabus shall be downloaded from the University Website (www.mangaloreuniversity.ac.in)


REGISTRAR

To,

1. The Chairman, Dept. of Biosciences, Mangalore University, Mangalagangothri
2. The Chairman, BOS in Biosciences, Dept. of Biosciences, Mangalore University.
3. The Registrar (Evaluation), Mangalore University.
4. The Superintendent (ACC), O/o the Registrar, Mangalore University.
5. The Asst. Registrar (ACC), O/o the Registrar, Mangalore University.
6. Guard File.

MANGALORE UNIVERSITY
DEPARTMENT OF BIOSCIENCES
SCHEME and SYLLABUS for TWO YEAR (FOUR SEMESTERS) M.Sc. in
BIOSCIENCES POST GRADUATE DEGREE PROGRAM UNDER CHOICE BASED
CREDIT SYSTEM (CBCS)

Preamble:

Based on directions of the University Grants Commission, New Delhi and Karnataka State Higher Education Council, the Choice Based Credit System (CBCS Semester Scheme) has been implemented. Mangalore University directed the Board of Studies (BoS) to frame the syllabus as per the regulations governing the Choice Based Credit System for the Two Year (Four Semester) Post-Graduate Programme. Accordingly, a syllabus approved by the BoS was in place since 2016.

This syllabus has now been revised keeping in mind the recent advancements in the field of Biological Sciences, the knowledge- and skill-based profile expected from a Master's in Biosciences along with fulfilling the requirement for the students' career prospects and was duly approved by the BoS in 2020.

The present M.Sc. in Biosciences Programme under CBCS Scheme has a total of 88 credits with 52 (59.09%) credits from Hard Core courses, 30 (34.09%) credits from Soft Core courses and 06 (6.97%) credits from Open Electives.

Programme Outcomes (PO)

PO1. Enhancement of state-of-the-art knowledge: Upgrade knowledge to develop general competencies and analytical skills on an advanced level required for teaching, research, industry, entrepreneurship and public administration in the field of biological sciences.

PO2. Skill-based use of tools and techniques: Independently operate various tools and acquire skills for the application of appropriate techniques to assess samples and carry out innovative studies on basic or applied aspects of biology.

PO3. Social Responsibility: Apply the knowledge of life sciences to contextually address specific issues in society with special reference to health and environment for well-being and sustainable development.

PO4. Effective Communication: Effectively communicate on diverse aspects of biology through oral presentations, written proposals, dissertations, reports, data analysis, interpretation and documentation.

Programme Specific Outcomes (PSO)

PSO1. Gain basic to advanced level knowledge in various branches of life sciences thus enabling students to build the confidence to pursue careers in academics, industries or become entrepreneurs in India and abroad.

PSO2. Empower with skill-based expertise and technical know-how in the field of biological sciences.

PSO3. Develop good communication skills with sound technical background in biological sciences, thus providing a strong foundation for both academic and industrial placements as well as setting up entrepreneurial ventures.

PSO4. Evolve in-depth scientific knowledge in various branches of biology.

PSO4. Explore, analyse and interpret lab- and field-based data using state-of-the-art techniques and tools in planning and executing innovative projects in life sciences.

M.Sc. BIOSCIENCES - SCHEME

I SEMESTER	Hrs/week	Credits
HARD CORE COURSES - THEORY		
BSH401 Biochemistry	4	4
BSH402 Cell Biology	4	4
BSH403 Basic Microbiology	4	4
SOFT CORE COURSES - THEORY (Any ONE to be opted)		
BSS404 Genetics	3	3
BSS405 Biochemical Techniques	3	3
PRACTICAL COURSES		
BSP406 Biochemistry Lab	4	2
BSP407 Cell Biology Lab	4	2
BSP408 Basic Microbiology Lab	4	2
BSP409 Genetics Lab	4	2
BSP410 Biochemical Techniques Lab	4	2
II SEMESTER	Hrs/week	Credits
HARD CORE COURSES - THEORY		
BSH451 Molecular Biology	4	4
BSH452 Biostatistics and Bioinformatics	4	4
SOFT CORE COURSES - THEORY (Any ONE to be opted)		
BSS453 Applied Microbiology	3	3
BSS454 Aquatic Biology	3	3
BSS455 Metabolism and Bioenergetics	3	3
PRACTICAL COURSES		
BSP456 Molecular Biology Lab	4	2
BSP457 Biostatistics and Bioinformatics Lab	4	2
BSP458 Applied Microbiology Lab	4	2
BSP459 Aquatic Biology Lab	4	2
BSP460 Metabolism and Bioenergetics Lab	4	2
OPEN ELECTIVE COURSES (Any ONE to be opted)		
BSE461 Biodiversity and Conservation	3	3
BSE462 Cancer Biology	3	3
III SEMESTER	Hrs/week	Credits
BSH501 Animal Physiology	4	4
BSH502 Plant Physiology	4	4
SOFT CORE COURSES - THEORY (Any TWO to be opted)		
BSS503 Applied Ecology	3	3
BSS504 Immunology	3	3
BSS505 Ecotoxicology	3	3
PRACTICAL COURSES		
BSP506 Animal Physiology Lab	4	2
BSP507 Plant Physiology Lab	4	2
BSP508 Applied Ecology Lab	4	2
BSP509 Immunology Lab	4	2
BSP510 Ecotoxicology Lab	4	2
OPEN ELECTIVE COURSES (Any ONE to be opted)		
BSE511 Pollution and Bioremediation	3	3
BSE512 Stem Cell Biology and Regenerative Medicine	3	3
BSE513 Behavioural biology	3	3
IV SEMESTER	Hrs/week	Credits
HARD CORE COURSES - THEORY		
BSH551 Biotechnology	4	4
SOFT CORE COURSES - THEORY (Any TWO to be opted)		
BSS552 Environmental Physiology	3	3
BSS553 Developmental Biology	3	3
PRACTICAL COURSES		
BSP554 Biotechnology Lab	4	2
BSP555 Environmental Physiology Lab	4	2
BSP556 Developmental Biology Lab	4	2
PROJECT WORK		
BSP557 Project Work (Report/Dissertation)	4	4

**M.Sc. BIOSCIENCES
(CBCS)
Scheme**

I SEMESTER

Code	Title	Teaching Hrs/week	Exam Hrs	Marks Exams	Marks IA	Total Marks	Credits
HARD CORE COURSES - THEORY							
BSH401	Biochemistry	4	3	70	30	100	4
BSH402	Cell Biology	4	3	70	30	100	4
BSH403	Basic Microbiology	4	3	70	30	100	4
SOFT CORE COURSES – THEORY (Any ONE to be opted)							
BSS404	Genetics	3	3	70	30	100	3
BSS405	Biochemical Techniques	3	3	70	30		
PRACTICAL COURSES							
BSP 406	Biochemistry Lab	4	3	35	15	50	2
BSP 407	Cell BiologyLab	4	3	35	15	50	2
BSP 408	Basic MicrobiologyLab	4	3	35	15	50	2
BSP 409	Genetics Lab	4	3	35	15	50	2
BSP 410	BiochemicalTechniques Lab	4	3	35	15		
Total						600	23

II SEMESTER

Code	Title	Teaching Hrs/week	Exam Hrs	Marks Exams	Marks IA	Total Marks	Credits
HARD CORE COURSES - THEORY							
BSH451	Molecular Biology	4	3	70	30	100	4
BSH452	Biostatisticsand Bioinformatics	4	3	70	30	100	4
SOFT CORE COURSES – THEORY (Any TWO to be opted)							
BSS453	Applied Microbiology	3	3	70	30	100	3
BSS454	Aquatic Biology	3	3	70	30		
BSS455	Metabolism and Bioenergetics	3	3	70	30		
PRACTICAL COURSES							
BSP 456	Molecular Biology Lab	4	3	35	15	50	2
BSP 457	Biostatistics and Bioinformatics Lab	4	3	35	15	50	2
BSP 458	Applied Microbiology Lab	4	3	35	15	50	2
BSP 459	Aquatic Biology Lab	4	3	35	15		
BSP 460	Metabolism and Bioenergetics Lab	4	3	35	15		
OPEN ELECTIVE COURSES (Any ONE to be opted)							
BSE461	Biodiversity and Conservation	3	3	70	30	100	3
BSE462	Cancer Biology	3	3	70	30		
Total						700	25

III SEMESTER

Code	Title	Teaching Hrs/week	Exam Hrs	Marks Exams	MarksIA	Total Marks	Credits
HARD CORE COURSES - THEORY							
BSH501	Animal Physiology	4	3	70	30	100	4
BSH502	Plant Physiology	4	3	70	30	100	4
SOFT CORE COURSES – THEORY (Any TWO to be opted)							
BSS503	Applied Ecology	3	3	70	30	100	3
BSS504	Immunology	3	3	70	30		3
BSS505	Ecotoxicology	3	3	70	30		
PRACTICAL COURSES							
BSP 506	Animal Physiology Lab	4	3	35	15	50	2
BSP 507	Plant PhysiologyLab	4	3	35	15	50	2
BSP 508	Applied Ecology Lab	4	3	35	15	50	2
BSP 509	Immunology Lab	4	3	35	15		2
BSP 510	EcotoxicologyLab	4	3	35	15		
OPEN ELECTIVE COURSES (Any ONE to be opted)							
BSE 511	Pollution and Bioremediation	3	3	70	30	100	3
BSE 512	Stem Cell Biology andRegenerative Medicine	3	3	70	30		
BSE 513	Behavioural Biology	3	3	70	30		
Total						700	25

IV SEMESTER

Code	Title	Teaching Hrs/week	Exam Hrs	Marks Exams	Marks IA	Total Marks	Credits
HARD CORE COURSES - THEORY							
BSH 551	Biotechnology	4	3	70	30	100	4
SOFT CORE COURSES - THEORY (Any ONE to be opted)							
BSS 552	EnvironmentalPhysiology	3	3	70	30	100	3
BSS 553	Developmental Biology	3	3	70	30		
PRACTICAL COURSES							
BSP 555	Biotechnology Lab	4	3	35	15	50	2
BSP 556	EnvironmentalPhysiology Lab	4	3	35	15	50	2
BSP 557	Developmental Biology Lab	4	3	35	15		
PROJECT WORK							
BSP 558	Project Work (Report/Dissertation)	-	-	70	30	100	4
Total						400	15
Grand Total						2400	82 + 6*

IA = Internal Assessment; * Not included for CGPA

Total Credits: 88(82+6*)

Hard Core credit: 18 + 12 + 12 + 06 + 04 (Project) = 52 (59.1%)

Soft Core credit: 05 + 10 + 10 + 05 = 30 (34.1%)

Open Elective credit: 03 + 03 = 06 (6.82%)

NOTE:

BASIS FOR INTERNAL ASSESSMENT: Internal Assessment marks in theory papers shall be awarded on the basis of theory test (70 Marks), Objective Test (15 Marks), Seminars and Assignments (15 Marks). The marks obtained shall be reduced to 30. Practical Internal Assessment marks shall be based on practical test and records. 30 marks for Practical Test and 05 marks for Class Records. The marks obtained shall be reduced to 15. 30 marks for Project Work (Report/Dissertation and Presentation/Viva).

THEORY QUESTION PAPER PATTERN: Question Papers in all the four semesters consists of three sections (Model question paper enclosed). Section I: Write short notes on any four out of six: (4x4=16 Marks) Section II: Write explanatory notes on any five out of seven: (5x6=30 Marks). Section III: Write any essay on any two out of three: (2x12=24 Marks). Questions are to be drawn from all the units of the syllabus by giving equal weightage to all the units.

PRACTICAL QUESTION PAPER PATTERN: 30 marks for practical exam proper (Major experiment - 10 marks, Minor experiments - 5x2=10 marks, Identify and Comment- 5x2=10marks) and 10 marks for Class Record.

PROJECT WORK may be conducted either in the Department or any other Institution or in an Industry. Project Report/Dissertation carries 70 marks and is evaluated as per regulations.

Model Question Paper
First Semester M.Sc. BIOSCIENCES Degree Theory Examination(CBCS)

Time:3 Hours

Max. Marks:70

Write short notes on **any four** of the following (not exceeding **2**pages **each**):(**4x4=16**)

- 1.a)
- b)
- c)
- d)
- e)
- f)

Write explanatory notes on **any five** of the following (not exceeding **3** pages **each**): (**5x6=30**)

- 2.
- 3.
- 4.
- 5.
- 6.
- 7.
- 8.

Write essayson**any two** of the following (not exceeding **8**pages **each**): (**2x12=24**)

- 9.
- 10.
- 11.

M.Sc. BIOSCIENCES (CBCS Semester Scheme) SYLLABUS
I SEMESTER
HARD CORE COURSES
BSH401 BIOCHEMISTRY

Course Outcomes:

Upon successful completion of the course, students will be able to:

- CO 1. Have in-depth knowledge of biochemistry and appreciate the knowledge of biochemistry in the day-to-day life
- CO 2. Demonstrate an understanding of basic biochemical principles with reference to structure and functions of proteins, carbohydrates and lipids, and their metabolic pathways.
- CO 3. Understand the mechanisms of transport and excretion of cholesterol and sterols
- CO 4. Know the clinical relevance of studying biomolecules and metabolic disorders.

Unit I (13 hours)

Carbohydrates: Classification, chemistry and properties of monosaccharides - Pentoses, hexoses, deoxyglucose amino sugars, muramic acid, neuraminic acid, disaccharides - Linkage in sucrose, lactose and maltose, polysaccharides - Homo- and hetero-poly saccharides - starch, cellulose, glycogen, hyaluronic acid, chondroitin sulphate, chitin, xylans, bacterial cell wall and blood group polysaccharides, glycoproteins. Metabolism of carbohydrates: Pathways and regulation. Glycogenesis and Glycogenolysis. Anaerobic glycolysis, Citric acid cycle, Hexose monophosphate shunt. Gluconeogenesis. Coordinated control of metabolism.

Unit II (13 hours)

Amino acids and Proteins: Classification, chemistry and properties of amino acids and proteins. Primary, secondary (alpha helix, beta pleated sheets), tertiary (fibrous - Collagen, globular - Myoglobin) and domain structure of proteins. Reverse turn and Ramachandran plot. Helix - coil, transition. Quaternary structure - Hemoglobin. Energy terms in biopolymers. Conformational calculations, hydrogen bonding, hydrophobic, electrostatic and Vander Waals interactions. Lipoprotein metabolism and associated disorders.

Unit III (13 hours)

Lipids. Classification, chemistry and properties of lipids. Biological role of phospholipids, Sphingolipids, Glycolipids and Plasmalogens. Structure of cholesterol, Structure and function of essential fatty acids, Eicosanoids, Prostaglandins, Thromboxanes, Leukotrienes. Metabolism of lipids. Biosynthesis of fatty acids, oxidation of fat and fatty acids - beta, alpha and Omega oxidation. Ketogenesis and ketolysis. Biosynthesis of phospholipids. Triacylglycerol biosynthesis and role of adipose tissues. Biosynthesis, transport and excretion of cholesterol and sterols.

Unit IV (13 hours)

Protein and amino acid metabolism. Nitrogen balance, transamination and deamination. Catabolism of phenylalanine, tyrosine, tryptophan, sulphur containing amino acids, creatine and creatinine. Urea cycle and disorders.

References:

1. Nelson, D. L., Cox, M. M. (2014). Lehninger Principles of Biochemistry, W.H. Freeman, New York.
2. Berg J.M., Tymoczko J.L., Stryer, L. (2010) Biochemistry, 6th Ed., W.H. Freeman, New York.
3. Zubay, G. (1998) Biochemistry, 4th Ed., WBC/McGrawHill.

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9. Buchanan B. B., Gruissem, W., Jones, R. L. (2005) Biochemistry and Molecular Biology of Plants, Courier Companies Inc
10. Skooge, A., Holler F. J., Nieman T. A. (2006) Principles of Instrumental Analysis, 6th Ed., Brooks/Cole
11. Voet, D., Voet, J. G., Pratt, C. W. (2006) Fundamentals of Biochemistry - Life at the Molecular Level, 2nd Ed., Wiley.
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14. van Holde, K. E., Johnson, W. C., Ho, P. S. (1998) Principles of Physical Biochemistry, Prentice Hall.
15. Harvey, R.A., Ferrier D. R., Champe P.C. (2007) Biochemistry, 4th Ed., Lippincott Williams and Wilkins
16. Satyanarayana U., Chakrapani U. (2008) Biochemistry, 3rd Ed., Elsevier Publishers
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19. Hames, D., Hooper, N. (2005). Biochemistry (BIOS Instant Notes). Taylor & Francis.

BSH402 CELL BIOLOGY

Course Outcomes:

Upon successful completion of the course, students will be able to:

- CO 1. Know the ultrastructural organization and functions of organelles of prokaryotes and eukaryotes.
- CO 2. Comprehend the general structure and molecular organization of chromosomes.
- CO 3. Gain theoretical knowledge how to use basic tools and techniques such as microscopy, centrifugation, autoradiography and centrifugation
- CO 4. Explain the physiochemical properties of biological membranes with structural and functional insights.
- CO 5. Understand the components of cell cycle control, mechanisms of cell division, apoptosis and senescence.
- CO 6. Understand how cells communicate one another and role of various messenger molecules in signal transduction.

Unit I (13 hours)

Ultrastructure of prokaryotic and eukaryotic cells: Ultrastructure, organization and functions of cell organelles: Endoplasmic reticulum, liposomes, Golgi complex and protein sorting, ribosomes and nucleus; Structure of mitochondrion, chloroplast - their genetic organization and their semiautonomous nature. Secretory and endocytotic pathway. Cytoskeleton-microtubules, microfilaments, intermediary filaments. Centriole, cilia, flagella and cell motility. Eukaryotic chromosome - Ultrastructure and molecular organization. Nucleosome model of chromatin structure, Heterochromatin and Euchromatin, Ultrastructure of Giant chromosomes, Structure and function of centromere and telomere. Microscopy: principles and applications of Light, Phase contrast, fluorescence, laser confocal, scanning and transmission electron microscopy. Autoradiography, cytophotometry and flow cytometry and centrifugation. Cytochemical and histochemical staining techniques.

Unit II (13 hours)

RBC as a Model membrane. Various models for membrane structure; Singer and Nicolson's model. Physicochemical properties of biological membranes – compositions, molecular organization, Membrane asymmetry – lipids, proteins and carbohydrates, lateral diffusion, membrane domains – caveolae, rafts. Transport across biomembranes- Energetics of membrane transport, Donnan membrane equilibrium, simple diffusion, osmosis, facilitated diffusion and active transport. Carrier proteins, Ion channels (voltage- and ligand-gated), Bacterial K^+ leak channel & aquaporin channels. Electrical properties of membranes- Membrane potential, Mechanisms of nerve conduction. Transmission across electrical and chemical synapse. Mechanisms of endocytosis and exocytosis.

Unit III (13 hours)

Components in cell cycle control - Cyclins, CDKs in yeast and mammalian cells. Check points in cell cycle. Mechanics of Cell Division- Different stages of mitosis. Cohesins and Condensins in chromosome segregation, Microtubules in spindle assembly, Structure of kinetochore, centrosome and its functions, Sister Chromatid separation. Cytokinesis role of actin & myosin in the generation of contractile ring. Meiosis – Significance. Chiasma formation - Synaptonemal complex. Recombination during meiosis - recombination nodules. Apoptosis: Mechanisms by internal signals and external signals, factors affecting apoptosis. Cell senescence.

Unit IV (13 hours)

Various types of cell signaling-endocrine, paracrine, juxtacrine and autocrine; Signaling molecules – hormones, neurotransmitters, gases, lipids, peptides. Overview of receptors: types (membrane and intracellular receptors), structure and regulation - G-protein coupled receptors, Ion channel receptors, Tyrosine kinase linked receptors & Receptors with intrinsic enzyme activity (RTK) and nuclear receptors. General mechanisms of signal transduction by G protein coupled receptors and receptor tyrosine kinase, Second messengers- Ca^{2+} , IP_3 , DAG, cAMP & cGMP – cellular effects. Signaling pathways in development and differentiation (overview). Cell-cell adhesion, cell junctions; Extracellular matrix, extracellular matrix receptors. Cell-cell and Cell-matrix interaction (Integrins and selectins and their interaction).

References:

1. Lodish, H., Berk, A., Zipursky, S.L., Matsudaira, P., Baltimore, D. (2007) Molecular cell biology, 6th Ed., WH. Freeman and company, New York.
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3. Kleinsmith, L. J. & Kish, V.M. (1995). Principles of Cell and Molecular Biology, Mc Laughlin, S., Trost, K., Mac Elree, E. (ed.), 2nd Ed., Harper Collins Publishers, New York.
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BSH403 BASIC MICROBIOLOGY

Course Outcomes:

Upon successful completion of the course, students will be able to:

- CO 1. Understand basic concepts, historical perspectives and contributions in Microbiology.
- CO 2. Understand evolution of prokaryotic and eukaryotic metabolism
- CO 3. Learn about microbial nutrition and culture of microbes in the laboratory.
- CO 4. Discern various factors affecting growth and death of microorganisms.
- CO 5. Explain the microbial metabolic pathways with their applications.

UNIT I (13 hrs)

Introduction to microbiology, historical perspectives, contributions of early microbiologists, Koch Postulates. Branches and scope of microbiology. Origin and evolution of microorganisms, discovery of anaerobic life, evolutionary chronology, trends in evolution of archaeobacteria, eubacteria and eukaryotes. Evolution of prokaryotic and eukaryotic metabolism. Modern methods of tracing and analysis of evolution.

UNIT II (13 hrs)

Microbial diversity, habitats, life cycles, structure and classification of bacteria, cyanobacteria, actinomycetes, fungi and viruses. Pathogenic microorganisms: bacteria, mycoplasmas, rickettsias, chlamydias and protozoa.

Microbial nutrition and cultivation: Nutritional categories of microorganisms, role of microbial nutrients; cultivation of aerobes, anaerobes and facultatives, obligate pathogens and viruses. Selective media, selective isolation and methods of preservation of microbes.

UNIT III (13 hrs)

Microbial growth, population and growth curves, generation time, batch and continuous cultures (e.g. chemostat, turbidostat), measurement of growth, microbiological assays (e.g. antibiotics, amino acids and vitamins).

Factors affecting growth and death of microorganisms: temperature, pH, water activity, O-R potential, salinity, hydrostatic pressure, disinfectants, antiseptics and chemotherapeutic agents. Methods of sterilization.

UNIT IV (13 hrs)

Microbial metabolism: Energy sources and classification; metabolism in autotrophs, heterotrophs; hexose and pentose phosphate pathways; synthesis of peptidoglycan, intermediary metabolism and secondary metabolites. Aerobic and anaerobic respiration, fermentation, electron transport system and substrate phosphorylation.

References:

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22. <http://www.microbiologymaven.com/>
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SOFT CORE COURSES

BSS404 GENETICS

Course Outcomes:

Upon successful completion of the course, students will be able to:

- CO 1. Gain in-depth knowledge in Genetics
- CO 2. Understand principles governing the inheritance and variations
- CO 3. Comprehend recombination in bacteria and development of rDNA technology.
- CO 4. Understand the phenomenon of mutation and learn skills to detect mutations

Unit I (13 hours)

Historical perspectives and scope of Genetics; Principles of Mendelian inheritance; Modifications of Mendelian monohybrid and dihybrid ratios-Incomplete dominance, Codominance, Lethal genes and Multiple alleles. Applications of Mendel's principles- the punnet square method, forked-line method, probability method; Formulating and testing genetic hypothesis-the chi-square-test, linkage and crossing over. Cytological basis of inheritance: Linkage and crossing over; Genetic mapping of chromosomes. Sex determination, Dosage compensation in mammals and drosophila. Sex linked inheritance (*Drosophila* and Human). Sex related traits, genetic disorders.

Unit II (13 hours)

Genetics of Bacteria: Transformation, transduction, Conjugation - Plasmids. Extra chromosomal inheritance with examples; Genomic organization in prokaryotes and eukaryotes; Laws of DNA constancy and C - value paradox. Mutations: Classification, types of mutations-deletion, duplication, translocation and inversion, spontaneous and induced mutations, molecular mechanisms of mutations. Biochemical basis for mutations; Detection of mutations – mutagenicity testing - Ames test, tests in drosophila (DLT, ClB, SLRL, SMART, ARLT) and mouse (DLT, MNT, Mitotic and meiotic, specific locus test, HMA)

Unit III (13 hours)

Genetic recombination at Molecular level: Reciprocal recombination, site specific recombination, models of recombination (Holliday model), Role of Rec A in Recombination. Transposable genetic elements: Bacterial transposons, Is elements, Composite transposons, Tn3 elements, Eukaryotic transposons-Ac and Ds elements in maize; P elements and Hybrid dysgenesis, Retrotransposons. Alu sequences. Human genetics: Human chromosomes, Chromosomal abnormalities-Sex chromosomal and autosomal; Genetic diseases, Pedigree analysis and genetic counseling, gene therapy.

References:

1. Gardner, E.J., Simmons M.J. & Snustad, D.P.(1991). Principles of Genetics. 8thEd. John Wiley and Sons, Inc., New York.
2. Hartl, D. L., Freifelder D. and Snyder, L.A.(1988). Basic Genetics. Jones and Bartlett Publishers, Boston.
3. Hollaender A. (Ed.). (1971-76). Chemical Mutagens. Principles and Methods for their Detection. Vols. 1, 2 & 3. Plenum Press, New York
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BSS405 BIOCHEMICAL TECHNIQUES

Course Outcomes:

Upon successful completion of the course, students will be able to:

- CO 1. Know the principle and applications of basic biochemical techniques.
- CO 2. Understand the role of biological solutions and calculations
- CO 3. Understand principle, instrumentation, applications and types of chromatography
- CO 4. Know the principle, instrumentation, applications and types of centrifugation
- CO 5. Understand the principle, instrumentation, applications and types of electrophoretic techniques

Unit I (13 hours)

Biological Solutions: preparation of solutions-Normality, molarity and molality: Acids and Bases, Buffers, salting in, salting out, Osmosis, Dialysis, Donnan Membrane Equilibrium, Viscosity of macromolecules, relationship with conformational changes, Density. **Chromatography** Principles of partition chromatography, paper, thin layer, column chromatography, ion exchange and affinity chromatography, gas chromatography, gel permeation chromatography, HPLC and FPLC.

Unit II (13 hours)

Centrifugation Principles of centrifugation, Svedberg's constant, concepts of RCF, different types of instruments and rotors, preparative, differential and density gradient centrifugation, analytical ultra-centrifugation, determination of molecular weights and other applications, subcellular fractionation. Filtration methods: Invention of filtration method. Various types of filter membranes and their applications.

Unit III (13hours)

Electrophoretic techniques Principles of electrophoretic separation. Continuous, zonal and capillary electrophoresis, different types of electrophoresis including paper, cellulose, acetate/nitrate and gel. Electroporation, pulse field gel electrophoresis, PAGE, SDS- PAGE and Iso electro focusing.

References:

1. Patabhi, V. & Gautham, N. (2003). Biophysics - NarosaPublHouse,
2. Khopkar, S. M. (2008). Basic Concepts of Analytical Chemistry, 3rd Ed., New Age Publications.
3. Upadhyay, A., Upadhyay, K., Nath, N. (2009). Biophysical Chemistry-Principles and Techniques, Himalaya Publ House
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9. Segal I. H. (1976) Biochemical calculation, 2ndEd.
10. Wilson, K. and Walker, J.(1996). Practical biochemistry.PrinciplesandTechniques. Cambridge Low PriceEditions
12. Shrikant, L. P. (2013) Understanding Biophysics. 4thEd., Suman Publications.
13. Krishna A. P. (2014) Text book of Medical Physiology, 2ndEd, Suman Publications.
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15. Gault, V. A., & McClenaghan, N. H. (2013).Understanding bioanalytical chemistry: principles and

- applications. John Wiley & Sons.
16. Van Emon, J. M. (Ed.). (2016).Immunoassay and other bioanalytical techniques. CRC Press
 17. Manz, A., Pamme, N., &Iossifidis, D. (2004).Bioanalytical chemistry. World Scientific Publishing Company.
 18. Ramesh, V. (Ed.). (2019).Biomolecular and Bioanalytical Techniques: Theory, Methodology and Applications. John Wiley & Sons.
 19. Hoppe, W., Lohmann, W., Markl, H., & Ziegler, H. (Eds.). (2012).Biophysics. Springer Science & Business Media.
 20. Jackson, M. B. (2006).Molecular and cellular biophysics. Cambridge University Press.

PRACTICAL COURSES

BSP406 BIOCHEMISTRY LAB

Course Outcomes:

After undergoing the course, students will be able to:

- CO 1. Develop skills required for biochemical qualitative and quantitative work
- CO 2. Learn methods to proteins, carbohydrates, lipids and NPN substances.
- CO 3. Operate instruments used in biochemistry labs
- CO 4. Conduct biochemical tests to diagnose some metabolic diseases.

1. Handling of pipette and understanding accuracy and precision of pipette
2. Qualitative analysis of carbohydrates: monosaccharides, disaccharides and polysaccharides
3. Qualitative tests for the proteins,
4. Qualitative tests for lipids and NPN substances.
5. Preparation of buffers and its pH determination
6. Preparation of normal, molar and percent solutions
7. Understand serial dilutions
8. Estimation of amino acids and nitrogen analysis by Micro-Kjeldahl method
9. Enzyme activity: Effect of temperature, pH, Km determination
10. Spectrophotometric estimation of metabolites: serum protein, sugar, creatinine, urea, uric acid
11. Colorimetric analysis of vitamins, ascorbic acid etc.,
12. Estimation of plant phenolics
13. Tests to measure glycosuria, proteinuria etc

BSP407 CELL BIOLOGY LAB

Course Outcomes:

After undergoing the course, students will be able to :

- CO 1. Acquire skills required in Cell Biology
- CO 2. Learn methods to study cell division and cell cycle
- CO 3. Develop skills in histological staining techniques isolate the sub-cellular organelles.
- CO 4. Perform experiments in cell biology

1. Micrometry and camera lucida drawings
2. Cell (RBC) counting using haemocytometer
3. Study of plasmolysis in cells of *Rheo* leaves.
4. Determination of mitotic index in onion root tips
5. Preparation of tissues for histology, Sectioning & Staining - Differential staining of tissue sections
6. Histochemistry-localization of a) Carbohydrates b) Proteins c) Nucleic acids
7. Hematoxylin staining and study on histology of liver, intestine, stomach, ovary, etc.,
8. Study of mitotic stages in onion root tip
9. Study of meiosis in Onion inflorescence/grasshopper testis
10. Study of chromosomal aberration in *Allium cepa* after chemical induction
11. Cell viability assays
12. Isolation of Sub cellular organelles
13. Measurement of Na-K ATPase in membrane fractions
14. Determination of osmotic fragility of erythrocyte membranes

BSP408 BASIC MICROBIOLOGY LAB.

Course Outcomes:

After undergoing the course, students will be able to :

- CO 1. Understand basic techniques and instrumentation in microbiology.
- CO 2. Apply the techniques of sterilization of media and glassware.
- CO 3. Isolate, identify and culture microorganisms
- CO 4. Perform microbial motility tests.
- CO 5. Execute the filter sterilization and microbial isolation.

1. Introduction to basic techniques and instrumentation in microbiology
2. Microscopic observations of microorganisms and micrometry
3. Staining techniques: Properties of stains, microbial smear preparation, simple and differential staining for morphological studies, Gram's staining, endospore staining, intracellular lipids, acid-fast staining, flagella, viability tests and relief (negative) staining;
4. Microbial motility tests.
5. Microbial culture media, microbial growth
6. sterilization of media and glassware, filter sterilization
7. stock culture, subculture, maintenance of culture.
8. Techniques of microbial isolation.

BSP409 GENETICS LAB

Course Outcomes:

After undergoing the course, students will be able to:

- Understand the importance of *D. melanogaster* as an excellent model in Genetics.
- Maintain and conduct experiments using *D. melanogaster*.
- Conduct crossing experiments to learn Mendelian and non-Mendelian Genetics
- Solve genetic problems such as legal issues like paternity and maternity disputes.

1. Salient features and method of maintenance of *Drosophila melanogaster* culture.
2. Techniques for handling and examining the flies.
3. Preparation of salivary gland chromosomes of *D. melanogaster* and identification of different arms.
4. Preparation of salivary gland chromosomes in *D. nasuta*
5. Identification of blood types in human.
6. Experiments to demonstrate patterns of inheritance of a few characters (Crossing).
7. Study of (i) mating behaviour in *Drosophila* (ii) somatic mitosis in *Drosophila*.
8. Biochemical separation of eye pigments in *Drosophila*
9. Genetic problems.

BSP410 BIOCHEMICAL TECHNIQUES LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Separate the mixtures by planar and column chromatographic techniques.
- CO 2. Undertake quality analyses required in food industry by identifying additives, vitamins, preservatives, proteins, sugars and amino acids.
- CO 3. Use UV-Vis spectrophotometry for estimation.
- CO 4. Operate flame photometry.
- CO 5. Perform electrophoretic techniques for separation and determination of molecular weight.
- CO 6. Perform immune-diffusion techniques and ELISA for detecting presence and

quantity of antigens.

CO 7. Use centrifugation for separation of molecules.

1. Ascending, descending and circular paper chromatography for separation of amino acids/carbohydrates
2. TLC of amino acids (1D and 2D)/carbohydrates
3. UV-Visible Spectrophotometry-verification of Beer Lambert's law
4. Flame photometry and its application in the estimation of serum, calcium, potassium and lithium and sodium.
5. HPLC (Demonstration)
6. Gel electrophoresis- native and SDS-PAGE and estimation of molecular weight of Proteins
7. ELISA for quantification of an antigen.
8. Immunodiffusion
9. Centrifuge use and application of centrifugation techniques for separation
10. Separation by filtration technology

II SEMESTER

HARD CORE COURSES BSH451 MOLECULAR BIOLOGY

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Understand the molecular basis of life.
- CO 2. Know the role of various enzymes involved in DNA replications.
- CO 3. Comprehend gene transcription and its regulation in prokaryotes and eukaryotes.
- CO 4. Understand protein synthesis and post-translational modifications
- CO 5. Understand the role of non-coding RNAs and miRNAs.

Unit I (13 hours)

Central dogma of molecular biology and its modification. Structure of nucleic acids; structure of DNA, topology, forms of DNA, repetitive DNA, DNA polymerases, DNA ligases, topoisomerases, gyrases, methylases, nucleases and restriction endonucleases, Ribonucleoproteins, Structure of m-RNA, Three dimensional structure of t- RNA, Heterochromatization, transposition, regulatory sequences and transacting factors, homologous recombination

Unit II (13 hours)

Organization of transcriptional units, Mechanism of DNA transcription in prokaryotes and eukaryotes, RNA processing (capping, polyadenylation, splicing, introns and exons), RNA polymerase, types, promoter initiation and transcription, DNA replication (Eukaryotes and prokaryotes) - Semi conservative, replication in *E.coli* and Eukaryote, control of replication, Replication in phage, plasmid and mitochondria, inhibitors of RNA synthesis and their mechanism of action, polycistronic and monocistronic RNAs, post transcriptional modification

Unit III (13 hours)

Protein synthesis in prokaryotes and eukaryotes, role of ribosomes and different types of RNA in protein synthesis, basic feature of genetic code - Triplet codon, Assignment of codons, degeneracy, variation in codon usage, universality, Amino acid activation, mechanism of initiation, elongation and termination, post translational modifications - Protein folding, role of chaperons. O and N glycosylation, Fatty acylation, attachment of glycosyl anchor, phosphorylation, other modifications, inhibitors of protein synthesis

Unit IV (13 hours)

Molecular basis of signal transduction in bacteria, plant and animals, Regulation of gene expression in bacteria and eukaryotes - fine structure of eukaryotic gene, exons, introns, repetitive DNA, Promoters enhancers, silencers, regulatory sequences, DNA-binding Proteins. Organization of Prokaryotic and eukaryotic genes, gene families, tandemly repeating genes, pseudogenes. **Operon Model:** Lac operon, catabolite repression. Negative and positive control, Trp operon attenuation, antitermination. Non-coding RNAs, microRNAs, Genome editing technologies.

References:

1. Alberts, B., Bray, D., Lewis, J., Raff, M., Roberts, K., Watson, J. D. (Eds.) (2007). Molecular biology of the cell. 5th Ed., Garland Publishing, Inc., New York.
2. Cooper, G.M. (2009) The cell-A molecular approach. 5th ed. Sunderland (MA), Sinauer

- Associates, Inc.
3. Gilbert, S.F. (2006) Developmental biology. 6thEd., Sunderland (MA), Sinauer Associates Inc.
 4. Lodish, H., Berk, A., Zipursky, S. L., Matsudaira, P., & Baltimore, D. (2007). Molecular cell biology, 6thEd., W.H. Freeman and company, New York.
 5. Karp, G. (2010). Cell and molecular biology-Concepts and experiments. 6thEd, John Harris, D. (ed.) Wiley & sons, New York.
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 9. Voet, D., Pratt, C.W., Voet J.G. (2008) Fundamentals of biochemistry: Life at the molecular level, 3rdEd. John Wiley & Sons

BSH452 BIOSTATISTICS AND BIOINFORMATICS

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Apply statistical methods to analyse and interpret the biological data.
- CO 2. Represent the data of experimental and field studies through graphs and diagrams.
- CO 3. Understand statistical concepts and learn to use a variety of statistical tests
- CO 4. Know how to use bioinformatics for DNA and protein sequence analysis through bioinformatics tools and databases
- CO 5. Understand microarray technique for gene expression analysis.
- CO 6. Understand the concept of protein folding and structure based targeted drug design

Unit I (13 hours)

Biological data-frequency distribution, graphical and diagrammatic representations; Measures of Central tendency - Mean, Median and Mode; Measure of Dispersion - Range, Variance, Standard deviation, Coefficient of variation, Diversity Index.

Populations versus sample - sampling techniques; Standard error, Confidence limits. Random experiment-probability. Binomial, poisson and Normal distributions and their applications in genetics.

Unit II (13 hours)

Simple linear Regression and Correlation analysis. Analysis of variance, principles of experimental design. Multiple regression.

Tests of significance- Normal, χ^2 , (Chi-square), 't' and F tests; Testing for goodness of fit. One-way analysis of variance (ANOVA) and Two-way analysis of variance. Statistical packages.

Unit III (13 hours)

Introduction to bioinformatics, databases, search engines, internet tools and World Wide Web (WWW). Molecular modeling database at NCBI, major web resources for bioinformatics - Biological database types and their functioning, microbiological databases, primary sequence databases, carbohydrate databases, RNA databases, genome databases, organism databases, biodiversity. Sequence database: Introduction, nucleotide sequence database, protein sequence databases, EMBL nucleotide sequence databases, structure databases. Phylogeny - Tree definitions, distance matrix methods and parsimony and bootstrapping. DNA and protein sequence Analysis, FASTA, BLAST and GCG Wisconsin/Emboss packages. Genomics and proteomics.

Unit IV (13 hours)

Microarray techniques, Gene Expression analysis, Protein Folding, Lattice models, Comparative modeling, threading, folds and function, Distributed Computing approach, genome@home, folding@home, proteomics, protein structure based targeted drug design – small molecular interactions and docking.

References:

1. Norman, T.J. and Bailey. (1981) Statistical methods in Biology. 2nd Ed. Hodder and Stoughton Ltd.
2. Arnold, E. (1979). Introductory statistics for Biology, 2nd Ed. London.
3. Campbell, R.C. (1983). Statistics for Biologists 2nd Ed. Cambridge Press.
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University Press,USA.

5. Sillince, J.A. and Sillince, M. (1991). Molecular databases for protein sequence and structure studies. Springer-Verlag
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7. Tisdall J. D. (2001) Beginning Perl for Bioinformatics, O'Reilly Press
8. Mount D. W. (2004) Bioinformatics: Sequence and Genome Analysis, CSHL Press
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10. Krane, D. E. & Raymer, M. L. (2002) Fundamental Concepts of Bioinformatics. Pearson.
11. Branden C. and Tooze J. (1991) Introduction to Protein Structure., Garland Pub.
12. Attwood, T. & Parry-Smith, D. (1999) Introduction to Bioinformatics., Pearson Ed.
13. Rosner, B. (2015). Fundamentals of biostatistics. Nelson Education.
14. Le, C.T., & Eberly, L.E. (2016). Introductory biostatistics. John Wiley & Sons.
15. Kaps, M., & Lamberson, W.R. (Eds.). (2017). Biostatistics for animal science. Cabi.
16. Forthofer, R. N., Lee, E. S., & Hernandez, M. (2006). Biostatistics: a guide to design, analysis and discovery. Elsevier.
17. Pevsner, J. (2015). Bioinformatics and functional genomics. John Wiley & Sons.
18. Xiong, J. (2006). Essential bioinformatics. Cambridge University Press.
19. Lesk, A. (2019). Introduction to bioinformatics. Oxford university press

SOFT CORE COURSES

BSS453 APPLIED MICROBIOLOGY

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Understand the applications of Microbiology in biomedical and industrial fields.
- CO 2. Comprehend the beneficial and harmful interactions of microbes with other organisms.
- CO 3. Use fermentation for production of ethanol, lactic acid and other industrial products.
- CO 4. Gain theoretical knowledge of food microbiology, prevention of air- and food-borne diseases and food poisoning.
- CO 5. Gain the basics of soil microbiology and its allied applications in agriculture.
- CO 6. Know the importance of aquatic microbiology and learn water purification and assessment of drinking water quality.

UNIT I (13 hrs)

Microbial Ecology: Microbial symbiosis, mutualism, plant-microbe interactions (e.g. mycorrhizas), animal-microbe interactions (human, ruminants and non-ruminants). Microbes in extreme environments - hydrothermal vents and coral reefs. Microorganisms as bio-indicators. Microbial bioremediation - role in environmental management, advantages and disadvantages. Ecological implications of genetically modified microorganisms. Fermentation: Ethanol, lactic acid, mixed acids, 2-3 butanediol, clostridial and propionic acid fermentation with emphasis on their ecological niches, merits and demerits.

UNIT II (13 hrs)

Food Microbiology: Classification of foods and oriental foods; Basic principles of food spoilage and methods of food preservation; Milk and milk products, milk microflora and their estimation, milk-borne diseases and prevention; Food poisoning, food-borne diseases and prevention.

Air Microbiology: Microflora of air and methods of their estimation, monitoring air allergens, air-borne diseases and prevention.

UNIT III (13 hrs)

Soil Microbiology: Soil microflora and methods of their estimation, role of soil microorganisms, bioconversion and decomposition. Biological nitrogen fixation (symbiotic and non-symbiotic), microbial phosphorus solubilization and their importance in soil fertility and agriculture.

Aquatic Microbiology: Microbes in water and wastewater and methods of their estimation (e.g. MPN), drinking water microbial standards and water purification; Water-borne diseases and prevention.

References:

1. Brock T.B. and Madigan M.T. (1991) Biology of microorganisms, Prentice Hall.
2. Pelczar J. and Chan E.C.S. (1981) Element of Microbiology, Mac Graw Hill, New York
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 - 11.Adams M.R and Moss M.O.(2003), Food Microbiology, 2nd Ed.,Panima Publ. Corp., NewDelhi
 - 12.Barrett J.T. (1998)Microbiology and Immunology Concepts, Lippincott-Raven,PA, USA
 - 13.Casida Jr., L.E. (1968) Industrial Microbiology. Wiley Eastern Ltd., New Delhi
 - 14.Elger, K.D. (2009) Immunology. John Wiley and Sons,USA
 - 15.Subba Rao N.S. (1982) Advances in Agricultural Microbiology. Oxford and IBH Pub., New Delhi.
 - 16.Arora, D.R. and Arora, B. 2012. Text Book of Microbiology, CBS Publ. & Dist. Pvt. Ltd., New Delhi.
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 - 18.Maier, R.M., Pepper, I.L. and Gerba, C.P. (2008) Environmental Microbiology, Academic Press
 - 19.Jjemba, P. K. (2004) Environmental Microbiology - Principles and Applications, Science Publ., USA

BSS454 AQUATIC BIOLOGY

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Gain theoretical knowledge in hydrobiology, abiotic factors and aquatic organisms.
- CO 2. Know how aquatic organisms adapted during the course of evolution.
- CO 3. Comprehend the importance of estuaries, mangroves, marshes, tidal flats, coastal wetlands and coral reef community.
- CO 4. Realize the impacts of aquatic pollution and how to use the biological strategies to prevent the pollution.
- CO 5. Know the basic concepts of biological productivity of both flora and fauna.
- CO 6. Gain the knowledge how to collect, separate and classify planktons, and their importance.
- CO 7. Appreciate the economic importance of hydrophytes and halophytes.

UNIT I (13 hrs)

Hydrobiology: Properties of water including sea water. Hydrological cycle. Ocean water movement - El nino effects. Structural and functional adaptations of aquatic organisms to the abiotic factors such as temperature, light, salinity, pressure and dissolved oxygen.

UNIT II (13 hrs)

Aquatic ecosystems: Freshwater habitats - wetland and swamps, tank/pond, river, lake/reservoir. Physico-chemical conditions and biological composition of estuaries, mangroves/marshes, tidal flats and coastal wetlands. Marine habitats - types of sea shore environmental parameters and adaptations of pelagic, benthic and deep sea organisms. Coral reef community. Aquatic pollution: characteristics, sources and types; eutrophication, red tide, shellfish poisoning; Biological control of aquatic pollution.

UNIT III (13 hrs)

Biological productivity: Basic concepts. Factors affecting productivity, measurement of productivity: Leibig's law of minimum, Shelford's law of tolerance. Production and distribution of aquatic fauna; Planktonology-classification, distribution, collection and separation of plankton; blooms/ swarms of plankton and algal production. Hydrophytes - types, adaptations, distributions and economic importance. Halophytes - types, adaptations, economic importance. Sea weed -types and their distribution and economic importance.

References:

1. APHA.(1992). Standard methods for examination of water and waste water. 19th Ed. APHA, New York, USA.
2. Edmondson, W.T. (1965). Freshwater Biology. John Wiley and Sons, New York.
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12. Raffaelli, D. and Hawkins, S. (1996). Intertidal ecology. Chapman & Hall, London. 356 pp.
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14. Davis, C.C. (1995). The marine and freshwater plankton. Michigan State University, Michigan. 502pp.

BSS455 METABOLISM AND BIOENERGETICS

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Understand the mechanisms and regulation of anabolic and catabolic processes
- CO 2. Know various disorders associated with metabolic pathways.
- CO 3. Understand the physiological importance and metabolism of vitamins.
- CO 4. Comprehend the concept of bioenergetics and thermodynamic principles in biology.

Unit I (13 hours)

Overview of metabolism, Metabolism of carbohydrates, pathways and regulation, gluconeogenesis, glycogenolysis, anaerobic glycolysis, citric acid cycle, hexose monophosphate shunt. Metabolism of lipids, Biosynthesis of fatty acids, Oxidation of fat and fatty acids, beta, alpha and omega oxidation, ketogenesis and ketolysis, metabolisms of acylglycerols and sphingolipids, cholesterol synthesis, transport and excretion, lipoprotein metabolism

Unit II (13 hours)

Protein and amino acid metabolism, nitrogen balance, transamination and deamination, catabolisms of aromatic and sulphur containing amino acids, urea cycle and disorders, Metabolisms of purines and pyrimidines, metabolism and functions of fat soluble A, D, E and K and water soluble B complex (B1, B2, B3, B5, B6, B7, B9 and B12) & C vitamins

Unit III (13 hours)

Bioenergetics, Thermodynamic principles in biology, Concept of free energy. Energy rich bonds, Coupled reactions, Electron transport chain, oxidative phosphorylation, group transfer, Biological energy transducers, inhibitors of electron transport chain, uncouplers

References:

1. Voet, D., Voet, G. (1994). Biochemistry. 2nd Ed., John Wiley and Sons
2. Stryer, L. (2004). Biochemistry. 4th Edition
3. Harper Biochemistry. Lange publications. 26th ed.
4. Lehninger, A.L., Nelson, D.L., Cox M.M.(2001). Principles of Biochemistry. CBS Publications
5. Devlin, T. M. (2005) Text-book of Biochemistry with clinical correlations 2nd Ed.

PRACTICAL COURSES

BSP456 MOLECULAR BIOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Perform agarose gel electrophoresis and realize its applications in biological research.
- CO 2. Isolate plasmid DNA, genomic DNA and total RNA from bacteria and other sources and determine their purity
- CO 3. Execute restriction digestion and mapping of DNA.
- CO 4. Design primers and run the PCR reaction.
- CO 5. Become skilled in gel documentation instrument (Geldoc) and image development.

- 1. Agarose gel electrophoresis
- 2. Isolation of plasmid DNA from bacteria and its identification by electrophoresis
- 3. Isolation of genomic DNA from various sources and its identification
- 4. Restriction digestion and mapping of DNA
- 5. Isolation of total RNA from various sources and gel electrophoresis
- 6. Design of primers and PCR
- 7. Determination of DNA/RNA purity by UV-Visible spectrophotometry
- 8. Demonstration of gel documentation and imaging

BSP457 BIOSTATISTICS AND BIOINFORMATICS LAB

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Perform suitable statistical tests for evaluation of data
- CO 2. Make suitable graphical representations of data
- CO 3. Perform statistical tests - t test, F-test, ANOVA
- CO 4. Develop the skill to use search engines, internet tools and databases.
- CO 5. Gain the practical knowledge of restriction mapping and microarray techniques.

Biostatistics

- 1. Measurement of Central tendencies, mean, median, mode
- 2. Measures of dispersion range SD, CV & SE
- 3. Scatter plot, Simple Correlation & Regression, Multiple Correlations
- 4. Construction of frequency table
- 5. Theoretical distribution, Binomial, Poisson & normal
- 6. Statistical inference, normal, t test, chi-square & F test
- 7. Analysis of Variance

Bioinformatics

- 1. Introduction to bioinformatics
- 2. Basic feature of computers; flow charts and problems.
- 3. Search engines and internet tools.
- 4. Biological databases
- 5. Use of databases (e.g. BLAST, FASTA)
- 6. Restriction mapping
- 7. Micro array techniques
- 8. Search engines
- 9. Web lab viewer and Ras mols

BSP458 APPLIED MICROBIOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Carry out quantitative and qualitative assessments of microflora of soil, water and air.
- CO 2. Selectively isolate and identify microbes using morphological and biochemical tools.
- CO 3. Understand the symbiotic association of microorganisms through experiments.
- CO 4. Assess microbial quality of drinking water and milk.
- CO 5. Perform microbiological assays for antibiotics and amino acids.

- 1. Quantitative and qualitative assessment of microflora of soil, water and air by direct and indirect methods.
- 2. Selective isolation of microbes (bacteria, actinomycetes, yeasts and fungi)
- 3. Studies on symbiotic association of microorganisms (rhizobia, cyanobacteria and arbuscular mycorrhizae)
- 4. Simple and special morphological and biochemical tests for identification of bacteria, fungi
- 5. Assessment of microbial quality of drinking water and milk
- 6. Microbiological assays (antibiotics and amino acids)

BSP459 AQUATIC BIOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Perform qualitative analyses of water samples for various parameters.
- CO 2. Identify freshwater, marine and benthic organisms
- CO 3. Estimate the productivity of aquatic ecosystems.
- CO 4. Understand the food and feeding habits in fish.

- 1. Water quality parameters
- 2. Freshwater, marine and benthic organisms.
- 3. Preparation of temporary and permanent slides of plankton.
- 4. Estimation of productivity.
- 5. Hydrophytes, halophytes and seaweeds.
- 6. Food and feeding habits in fish.
- 7. Sewage organisms.
- 8. Instrumentation in aquatic biology and field trips

BSP460 METABOLISM AND BIOENERGETICS

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Perform spectrophotometric estimation of various metabolites.
- CO 2. Diagnose some of the metabolic diseases through biochemical tests.
- CO 3. Quantify vitamins and phenolics in plant samples.
- CO 4. Calculate standard free energy change, redox potential, and mitochondrial respiration.

- 1. Spectrophotometric estimation of metabolites: serum protein, sugar, creatinine, urea, uric acid
- 2. Colorimetric analysis of vitamins, ascorbic acid etc.,
- 3. Estimation of plant phenolics
- 4. Tests to measure glycosuria, proteinuria etc
- 5. Calculations in Bioenergetics: standard free energy change, redox potential, mitochondrial respiration etc

OPEN ELECTIVE COURSES

BSE461 BIODIVERSITY AND CONSERVATION

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Understand the relevance of biodiversity and conservation.
- CO 2. Describe the levels of biodiversity organizations.
- CO 3. Understand Indian ecological/geographical diversity, including Himalayan region, Desert, Western Ghats, Coastal region and Hotspots of biodiversity.
- CO 4. Understand microbial diversity and its importance.

Unit I (13 hours)

Basic concepts and definitions, scope, biosphere, habitats, food chain, food web. Levels of biodiversity organizations – Genetic diversity, Species diversity and Ecosystem diversity. Indian ecological/geographical diversity: Himalayan Region, Deserts, Semiarid region, Gangetic plains, Western Ghats, Coastal region; Hotspots of biodiversity, Microbial diversity: Bacteria, Cyanobacteria, Fungi and Lichens, Algae, Protozoa and viruses, habitat. Mushrooms – edible and nonedible. Plant and animal association with microbes. Beneficial and harmful microbes, Culture, Cultivation of bacteria. Microbial products.

Unit II (13 hours)

Plant diversity: Lower and higher group of plants, plant ecosystem and its classification. Major ecosystem types, tropical forests, temperate forests. Arid and Semiarid ecosystems, boreal forests, Arctic and Alpine systems, grasslands, wetland ecosystem. Marine ecosystems, Epiphytes, parasites and orchids. Values and uses of plant diversity. Animal diversity: Lower and higher group of animals, their ecological niches. Zoogeographical regions of the world and India. Animals in temperate, tropical and boreal forests, cave and mountains, Coastal ecosystems, mangrove and estuaries, coral reefs.

Unit III (13 hours)

Biodiversity Conservation: Causes and prevention of Plant and Animal biodiversity loss; IUCN Red List Categories and Criteria. Conservation strategies – *Ex-situ* and *In-situ* conservation, Protected ecosystems – Biosphere reserves, National parks, Sanctuaries, Botanical gardens, Sacred groves; Wildlife conservation and wildlife conservation act; Centers of diversity study.

References:

1. Daniel, J.C. A century of natural history. Bombay natural History Society, Bombay, 697pp.
2. Dwivedi, A.P. (1993). Forests. International book Distributors, Dehra Dun. 352pp.
3. Odum E. P. (1983). Basic Ecology. Saunders College, London.
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6. Korringa, P. (1976). Farming of marine organisms law in the food chain. Elsevier, Amsterdam. 264pp.
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8. Lieth, H. (1989). Tropical rain forest ecosystems. Elsevier, Amsterdam. 713pp.
9. Southwood, T.R.E. (1978). Ecological methods, Chapman and Hall, London. 524pp.
10. Tiwari, S.K. (1985). Readings in Indian Zoogeography. Today and Tomorrow's Printers and Publishers, New Delhi. 604pp.

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12. Reddy, P.A. (2000). *Wetland ecology*. Cambridge University Press, London. 614pp.
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14. Brummit, R.K. (1992). *Vascular Plant Families and Genera*, Royal Botanic Gardens, Kew, England.
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20. Kushalappa, C.G. and Bhagwat, S.A. (2001). Sacred groves: Biodiversity, threats and conservation. In: Uma Shanker, R., Ganeshaiah, K.N. and Bawa, K.S. (Eds.) *Forest genetic resources: Status, threats and conservation strategies*. Oxford & IBH Publishing Co. Pvt. Ltd., New Delhi, pp.21-29.
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26. Lkr, L. (2013). Indigenous techniques and practices for management of bio-resources: a Naga experience. *International journal of Bio-resource and Stress Management*, 4(4), 648-650.
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BSE462 CANCER BIOLOGY

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Understand the molecular biology and etiology of cancer.
- CO 2. Understand cell transformation mechanisms and role of oncogenes and tumoursuppressorgenes.
- CO 3. Perform the tests for identification of different types ofcancers.
- CO 4. Describe carcinogenic agents
- CO 5. Understand diagnosis and conventional and advanced cancer therapies.
- CO 6. Understand mechanisms of neoplasia and signalingpathways.

Unit I (13 hours)

Cellular hallmarks of cancer, Molecular biology of cancer development, Cell transformation mechanisms, benign and metastatic tumour, Protooncogenes, Oncogenes and tumour suppressor genes, Cellular senescence, Telomeres, cellular immortalization and tumorigenesis, Carcinogen- types and identification tests

Unit II (13 hours)

Multistep tumorigenesis, Mechanisms of neoplasia and signaling, tumor virology, Growth factors, receptors and cancer, cytoplasmic signaling circuitry programs and cancer, Cell cycle control, Genome integrity and cancer, pRb and control of cell cycle clock, DNA damage checkpoints and repair, Mismatch repair pathway and cancer.

Unit III (13 hours)

Tumor immunology and immunotherapy, rational design of Cancer therapeutics and diagnostics, Cancer nanotechnology, sequelae of cancer and its treatment, Genomic and proteomic technologies and application of new technologies in prevention, assessing risk, diagnostics and treatment of cancer.

References:

1. Weinberg R. A. (2006). Cancer Biology, GarlandScience.
2. Mckinnell R. G., Parchment R. E., Perantoni, A. O. and Pierce B. (1998), The Biological Basis of Cancer. Cambridge UniversityPress.
3. Kleinsmith, J. L. (2005), Principles of Cancer Biology, Benjamin CummingsPublication.
4. Franks L. M. and Teich N. M. (1997), Introduction to the Cellular and Molecular Biologyof Cancer (3rdEd.), Oxford UniversityPress.
5. Ruddon R. W. (2007). Cancer Biology, 4thEd. Oxford University Press
6. Roger John Benjamin King, Mike W. Robins, Cancer Biology, Pearson/Prentice Hall, 2006.
7. Pelengaris, S.& Khan, M. (Eds.). (2013).The molecular biology of cancer: A bridge from bench to bedside. John Wiley & Sons.
8. Pecorino, L. (2016).Molecular biology of cancer: mechanisms, targets, and therapeutics. Oxford University Press, USA.

**II SEMESTER
HARD CORECOURSES
BSH501ANIMAL PHYSIOLOGY**

52hrs

Course Outcomes:

After successful completion of the course, students will be able to :

- CO 1. Gain in-depth understanding of gastrointestinal system, associated disorders, digestive processes and mechanism of absorption of nutrients.
- CO 2. Comprehend ultrastructure and functioning of nerves and muscles.
- CO 3. Understand the importance of various endocrine glands, associated disorders, hormones and their mode of action
- CO 4. Understand osmoregulation and excretion mechanisms and modes across organisms.
- CO 5. Comprehend the concept of thermoregulation and adaptive features.
- CO 6. Develop in-depth understanding of sensory receptors

Unit I (13 hours)

Gastrointestinal System: Digestive processes and mechanisms of absorption of dietary carbohydrates, proteins and lipids; coordination of digestive and absorptive activities; gastrointestinal disorders.

Nervous system: Neuron and nerve impulse conduction synapses, synaptic transmission and neurotransmitters; reflex mechanisms; functions of the sensory and motor areas of the CNS; autonomic nervous system.

Unit II (13 hours)

Endocrine system: Hypothalamus. Endocrine glands - pituitary, thyroid, parathyroid, adrenals, pancreas, ovary, testis, pineal, GI tract and placenta: hormones - release, transport, mechanism of action and biological action; Neurohormones of the hypothalamus; endocrine disorders, Neuroendocrine system in Insecta and Crustacea.

Muscular system: Contraction of skeletal muscle; molecular basis of muscle contraction; energetics of muscular contraction; neuromuscular transmission and excitation contraction coupling; muscle atrophy and dystrophy.

Unit III (13 hours)

Osmoregulation and excretion: Biological significance of water; Osmoregulation in aquatic and terrestrial vertebrates; regulatory mechanisms; Major functions of excretory system; Organs of excretion- Basic processes responsible for the formation of the excreted fluid; Functional types- Generalized excretory organs and Specialized excretory organs; Classification of excretory organs and their distribution in the animal Kingdom; General patterns of nitrogen and non-protein nitrogen excretion; physiology of urine formation in mammals; renal diseases.

Unit IV (13 hours)

Thermoregulation: Thermoregulation-a phenomenon of homeostasis; Thermoregulatory adaptations-Physiological, Physical and Behavioral adaptations; Thermoregulation in aquatic and terrestrial invertebrates; Thermoregulation in Vertebrates-Fishes, Amphibians, Reptiles, Birds and Mammals.

Receptor system: Sensory receptors-classification and properties; Receptor Mechanisms: Chemoreceptors- gustatory receptors and olfactory receptors, Mechanoreceptors- Touch or pressure receptors, Pain receptors, Receptors concerned with equilibrium, gravity, acceleration and vibration, Phonoreceptors; Electromagnetic receptors- Photoreceptors

Thermoreceptors; Special Senses- Neurophysiology of Vision, Hearing and Chemical senses.

References:

1. Berne, R.M. & Levy, M.N. (1991). Physiology. The C.V. Mosby Company, St.Louis.
2. Ganong, W.F. (1999). Review of Medical Physiology (19th Ed) Kotheri Book Depot, Bombay.
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10. Rastogi, S. C. (2007). Essentials of animal physiology. New Age International.
11. Schmidt-Nielsen, K. (1997). Animal physiology: adaptation and environment. Cambridge University Press.
12. Schulte, P. M. (2013). Principles of Animal Physiology: Pearson New International Edition. Pearson Education Limited.
13. Sapolsky, R. M. (2017). Behave: The biology of humans at our best and worst. Penguin.
14. Kurpad, A., Vaz, M., & Raj, T. D. (2013). Guyton & Hall: Textbook of Medical Physiology-A South Asian Edition. Elsevier India

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Understand the role of various nutrients in plant growth
- CO 2. Comprehend the various concepts of water relation in plants and physiological processes.
- CO 3. Gain in-depth knowledge on photosynthesis and regulatory mechanisms.
- CO 4. Understand role of various growth regulators in plant growth
- CO 5. Gain knowledge on different methods and tools of plant breeding
- CO 6. Understand plant pathology

Unit I (13 hours)

Plant nutrition: Trace elements and their role, major and minor elements in soil and plants; Essentiality of elements- Sand culture, Soil culture, Hydroponics, Aeroponics; Mineral deficiencies and their rectification, nitrogen, phosphorus and sulfur metabolism.

Unit II (13 hours)

Water relations in plants: water requirements, Physical forces involved in water absorption, Osmotic system, Water potential, Site and path of water absorption; Ascent of Sap, mechanism of translocation of water and solutes; Factors affecting water absorption; Transpiration- Types of transpiration, structure and functions of stomata, mechanism of stomatal movement, Factors affecting transpiration, Guttation, anti-transpirants. Photosynthesis: Chloroplast and photosynthetic pigments; Concept of photosynthetic unit; Oxygenic and anoxygenic photosynthesis; Concept of pigment system; Stages of photosynthesis- cyclic and non-cyclic photophosphorylation; Hill reaction, Photorespiration; carbon dioxide fixation in C₃ and C₄ plants, CAM plants; Factors affecting photosynthesis.

Unit III (13 hours)

Plant Growth and Growth Regulators- Plant growth, Growth curve, measurement of growth, Phytohormones: Biosynthesis, Mechanism of action and application of auxins, gibberellins, cytokinins, ethylene, abscisic acid; Vernalin, Florigen, Morphactins; Phytochromes. Plant breeding: Objectives – high yield, improved quality, disease and pest resistance, early maturity, photosensitivity, varieties for new seasons, resistant varieties. Breeding in self-pollinated crops. Methods of breeding- Selection, Backcross method, Hybridization- objectives, types, procedure. Mutagenesis.

Unit IV (13 hours)

Plant pathology- Plant pathology in relation to important diseases of crop plants. Important plant diseases: Plant diseases caused by viruses, mycoplasma, bacteria, fungi, protozoa, nematodes, parasitic angiosperms - symptoms, etiology, life cycle, transmission etc. Seed borne diseases and transmission: Pollination, fertilization, embryogenesis, morphology and physiology in relation to seed infection. Seed-borne pathogens and their importance - viruses, bacteria, fungi and nematodes; seed infection and contamination.

References:

1. Hopkins, W.G. (1995). Introduction to Plant Physiology, John Wiley and Sons, Inc. New York.
2. Devlin, R.M. (1983). Plant Physiology. CBS Publications & Distributors, New Delhi.

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13. Sharma, J. R. (1994). Principles and practice of Plant Breeding. Tata McGraw Hill Publishing Co. Ltd. New Delhi. pp 599.
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SOFT CORE COURSES
BSS 503APPLIED ECOLOGY

39hrs

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Understand biodiversity, hotspots, conservation and management
- CO 2. Develop knowledge of forest and landscape ecology and watershed management.
- CO 3. Understand fisheries and aquaculture methods for commercial production of sea food
- CO 4. Learn about the impacts of aquatic pollution.
- CO 5. Develop in-depth knowledge in population ecology, prey-predatory dynamics, life-history strategies, energy budgets and reproductive strategies.

Unit I (13 hours)

Biodiversity: types, significance, distribution and measurements-Species richness: Simpson index, Shannon Wiener index, Evenness. Megadiversity countries, hot spots, biodiversity of Western Ghats and Eastern Himalayas. Wildlife management: Present status of threatened wildlife of Western Ghats; Conservation, Administrative and Judicial measures. Forest and landscape ecology: types of forests and their distribution with reference to Western Ghats; Vegetation mapping; Plant-animal interactions; Integrated pest management. Landscape Ecology – watershed management.

Unit II (13 hours)

Fisheries: Aquatic resources - fish, mollusca and crustaceans. Aquatic wildlife; Conservation and management of aquatic wildlife. Aquaculture - prawns, seaweeds, oysters, mussels, fin fishes and the environment. Aquaponics. Aquatic pollution – Eutrophication, algal blooms, coral reefs – bleaching, shellfish poisoning. Ganga action plan

Unit III (13 hours)

Population ecology: Demography-life tables; population structure-recruitment patterns, settlement and migration; population growth-growth patterns, age and growth, allometry, growth parameters; biotic parameters-predation, prey-predatory dynamics, competition, mutualism and population regulation; life history strategies-life history traits, longevity and survival rates, energy budgets, and reproductive strategies, *k*-selection and *r*-selection.

References:

1. Burn, A.J., Coaker, T.H. and Jepson, P.C. (1987). Integrated pest management. Academic Press, London. 474pp.
2. Daniel, J.C. A century of natural history. Bombay natural History Society, Bombay. 697 pp.
3. Dwivedi, A.P. (1993). Forests. International book Distributors, Dehra Dun. 352pp.
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22. Pielou, E.C., mathematical ecology. Wiley, New York, 385pp.
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24. Qasim, S.Z. (1999). The Indian Ocean Images and realities. Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi. 340 pp.
25. Roughgarden, J.J., 1987 Science of ecology. MacMillan, New York. 710pp.
26. Sivaraju, V.V., and Balachandran, I. (1994). Ayurvedic drugs and their plant sources. 27. Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi. 570pp.
27. Southwood, T.R.E. (1978). Ecological methods, Chapman and Hall, London. 524 pp.
28. Techniques to maintain biological diversities, 1988. SIRC, London.
29. Tiwari, S.K., (1985). Readings in Indian Zoogeography. Today and Tomorrow's Printers and Publishers, New Delhi. 604pp.
30. Zaika, V.E., (1970). Specific productivity of aquatic invertebrates. Wiley, New York, 154 pp.

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Gain an in-depth knowledge of immunology.
- CO 2. Understand the structure and functions of various immune cells and organs
- CO 3. Comprehend antigen and antibody structure and the mounting of immune responses.
- CO 4. Understand autoimmunity, hypersensitivity and immunodeficiencies
- CO 5. Understand the principles and application of various immunological techniques.

UNIT I (13 hrs)

Immunology: History and scope of immunology; Immunity, classification of immunity; Host defence: cellular, tissue and humoral immunity; Acquired immunity; Primary and secondary lymphoid organs; Immune response of T and B cells; Cytokines – structure and functions. Microbial defence: invasion, antigens, toxins. Antibodies: Production, structure, classification and functions; hyper variable region, Isotypic, allotypic and idiotypic variations. Antigenicity and immunogenicity, haptens. Complement.

UNIT II (13 hrs)

Autoimmune diseases: Thyrotoxicosis, Systemic Lupus Erythematosus, Antinuclear antibodies. Hypersensitivity - reactions. Tumor immunology– tumor antigens, immunosurveillance, immunological escape. Immune deficiency diseases– AIDS; Immunological tolerance, Immunization and Vaccines: Types and production.

UNIT III (13 hrs)

Major Histocompatibility Complex (MHC), HLA polymorphism. Tissue haplotypes and disorders, Tissue and organ grafting, graft rejection, Immune suppression. Immunological techniques: Antigen-antibody reactions. Precipitation and agglutination, immunodiagnosis, ELISA, RIA, immunoblotting and immunofluorescence and chemiluminescence; Fluorescent activated cell sorter (FACS); Hybridoma technology, production and application of monoclonal antibodies.

References:

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2. Ivan Riott (1988) Essential immunology –8th edition Blackwell publishers,
3. Wier DM (year) Handbook of expt. Immunology vol.1,2. Blackwell scientificPub.
4. Janis Kuby. (year) Immunology –Freeman and co publishers,2000
5. Ivan Riott, Jonathan Brostoff and David Male. (year) Immunology – 3rd edition. Mosbypublishers
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8. Elgert KD. (year) Immunology. Jon Wiley and Sons,USA
9. Barrett JT, (year) Microbiology and Immunology Concepts. Lippincott-Raven,USA

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Know the principle of bioassays for assessment of toxicity.
- CO 2. Understand how the biotransformation and detoxification of xenobiotics occurs
- CO 3. Gain the knowledge how to do the toxic risk and environmental impact assessments.
- CO 4. Understand various atmospheric toxicants and consequences of air pollution, acid rain, photochemical smog, global warming, ozone depletion and haze.
- CO 5. Gain in-depth knowledge of the adverse effects of alcohol, tobacco, food additives, petroleum and petroleum products
- CO 6. Understand the impact of pesticides and metal toxicity
- CO 7. Know antidote therapies for pesticide poisoning.

Unit I (13 hours)

Introduction, definition and various facets of ecotoxicology; Kinds of toxicity; time & dose-response relationships; factors influencing the toxicity; Bioassay- toxicity testing; Role of US-FDA. Metabolism of toxic substances: biomagnification, biotransformation and detoxification; Effects of environmental toxicants- sub cellular, cellular, individual, population and ecosystem levels. Toxic risk assessment: Methods, monitoring, importance and surveillance of risk assessment; Environmental Impact Assessment.

Unit II (13 hours)

Atmospheric toxicants: Major sources, types and standards; Primary pollutants- Carbon monoxide, sulphur oxides, nitrogen oxides, particulate matter, hydrocarbons, asbestos and CFCs; Secondary pollutants; Impact of air pollutants on climate-Acid rain, photochemical smog, global warming, ozone depletion and haze. Toxicity of Alcohol, tobacco & its products, food additives, petroleum & petroleum products.

Unit III (13 hours)

Pesticides: Definition, classification, usage and exposure; Insecticides: Organochlorines - DDT, cyclohexane, aldrin and endosulfan poisoning and treatment; Organophosphates and carbamates-Examples, sources, effects and treatment; herbicides, fungicides, rodenticides, endocrine disrupters. PCBs and Dioxins. Metal toxicity - History, sources, emissions, effect of mercury, cadmium, arsenic and lead on metabolism and environment. Poisoning - antidote.

References:

1. Boudou, A. (1997). Aquatic toxicology. Vol. I and II.
2. Diwakar Rao, P.L. (1990). Pollution control Hand book, Utility Publications Ltd., Secunderabad, India.
3. Eaton, A.D., Clesceri, L.S. & Greenberg, A.E. (1995). Standard Methods for the Examination of Water and Wastewater. APHA, Washington.
4. Gupi P.K. and Salunke, D.K. (1985). Modern Toxicology. Vol. I, II and III. Metropolitan Publications, Delhi.
5. Hommadi, A.H. (1990). Environmental and Industrial safety. Indian Bibliographics Bureau, Delhi.
6. Jorgensen, S.E., Modelling in Ecotoxicology. Elsevier, Amsterdam.
7. Lewin, S.A. et al., (1989). Ecotoxicology: Problems and approaches. Springer - Verlag, Tokyo, New York.
8. Moriarty, F. (1975). Pollutants and animals: A factual perspective. George Allan & Unwin

Ltd.,London

9. Omkar, (1995). Concepts of Toxicology. Chand & Co., Jalandhar.
10. Schmitz, R.J. (1996). Introduction to water pollution biology. Asian Books Pvt. Ltd., New Delhi.
11. Trivedi, P.R. and Sudarshan, K. (1995). Global environmental issues. Commonwealth Publications, New Delhi.
12. Vernberg et al. (1981). Biological monitoring of marine pollutants. Academic Press, New York

PRACTICAL COURSES

BSP 506 ANIMAL PHYSIOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Perform experiments to estimate enzyme activity and understand factors affecting enzyme activity
- CO 2. Perform experiments on hormonal control of reproductive biology.
- CO 3. Perform experiments in muscle physiology and osmoregulation.
- CO 4. Conduct qualitative tests for excretory products and demonstrate active transport

- 1. Gastrointestinal function–
 - 1.1. Factors affecting enzyme activities in digestion of foodstuffs.
 - 1.2 Quantitative estimation of Enzyme (amylase) activity.
- 2. Neuroendocrinology–
 - 2.1 Effect of hormones on blood glucose in rats.
 - 2.2 Study of estrous cycle in mice
 - 2.3 Study of sperm count, sperm morphology and sperm motility
- 3. Muscle Physiology–
 - 3.1 Histochemical detection of SDH activity in red and white muscle fibres.
- 4. Osmoregulation–
 - 4.1 Estimation of Fluid balance in an animal.
 - 4.2 Osmotic relationship in animals at the level of cell as well as entire organism.
- 5. Excretion–
 - 5.1 Qualitative tests for excretory products.
 - 5.2 Demonstration of active transport.

BSP507 PLANT PHYSIOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Realize the importance each nutrient in plant growth through experimentation and observation.
- CO 2. Observe mineral deficiency symptoms in plants.
- CO 3. Know how to perform the tests for understanding water relations.
- CO 4. Understand the photosynthesis by conducting some allied experiments.
- CO 5. Understand the role of growth hormones in plants.

- 1. Plant nutrition–
 - 1.1 Observation of mineral deficiency symptoms in plants.
- 2. Water relations–
 - 2.1 Experiments to demonstrate the diffusion pressure deficit in plant cell.
 - 2.2 Determination of stomatal index, stomatal frequency and measurement of stomatal aperture.
 - 2.3 Determination of water potential
- 3. Photosynthesis –
 - 3.1 Separation and estimation of chloroplast pigments.
 - 3.2 Demonstration of Kranz anatomy
- 4. Growth hormones and their regulation–
 - 4.1 Experiments to demonstrate the effect of hormones on shoot apex.
- 5. Plant pathology
 - 5.1 Pathogens in crop plants

BSP508 APPLIED ECOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Enhance the theoretical knowledge of applied ecology with lab experiments and field visits.
- CO 2. Understand plant-animal interactions and prey-predator relationship.
- CO 3. Unravel medicinal properties of plants and significance of conservation
- CO 4. Develop skills of remote sensing.
- CO 5. Identify the freshwater and marine fishery resources.
- CO 6. Estimate growth parameters and determine the probability of death.

- 1. Biodiversity
- 2. Terrestrial biodiversity
- 3. Aquatic biodiversity
- 4. Plant-animal interactions
- 5. Endangered medicinal plants.
- 6. Landscapes analysis through remote sensing data.
- 7. Freshwater fishery resources
- 8. Marine fishery resources
- 9. Estimation of growth parameters
- 10. Life-tables
- 11. Prey-predator relationships

BSP509 IMMUNOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Develop skills in immunology lab experiments.
- CO 2. Isolate lymphocytes and identify different blood cells
- CO 3. Understand hemolymph cells in insects
- CO 4. Perform immunoassays using various immunodiffusion methods
- CO 5. Detect and quantify antigens and allergens using established methods

- 1. Study of immune system in rats
- 2. Blood film preparation and study of immune cells
- 3. Isolation of lymphocytes
- 4. Study of insect hemocytes
- 5. Ouchterlony double diffusion assay
- 6. Radial Immunodiffusion technique
- 7. Immunological diagnosis of pregnancy and infection
- 8. DOT- ELISA technique
- 9. Rocket immunoelectrophoresis method
- 10. Detection of allergens: Pollen Count by sticky slide method

BSP 510 ECOTOXICOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Learn and practice safety measures to be taken in laboratories.
- CO 2. Determine acute and chronic toxicities through bioassays.
- CO 3. Estimate oil and grease from water and differentiate between clean and polluted water samples

CO 4. Perform tests for detection of metals and other toxic pollutants and food adulterants.

CO 5. Assess effect of metals on plant growth

1. Good Laboratory Practices
2. Safety notices in environmental toxicological studies.
3. Bioassay experiments using different test systems.
4. Behavioural study of the fish under exposure to toxicants.
5. Experiments on solid waste
6. Estimation of oil and grease in water sample.
7. Demonstration of catalase activity in polluted waters.
8. Spot test for detection of metals, residual chlorine, nitrite poisoning, fluoride toxicity, food adulterants and pesticide residues.
9. Effect of CdCl_2 on germination of Bengal gram.
10. Effect of toxicants in meristematic tissue (Onion root tips).
11. GC analysis of pesticide residues in food samples.

OPEN ELECTIVE COURSES
BSE 511 POLLUTION AND BIOREMEDIATION

39hrs

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Understand causes and effects of environmental pollution and bioremediation.
- CO 2. Know about air, water and land pollutants and their impact.
- CO 3. Realize the impacts of water pollution on aquatic biota and human health.
- CO 4. Know the causes of acid rain, photochemical smog, global warming, ozone depletion and haze.
- CO 5. Understand the concept of bioremediation and how to use microorganisms, plants and enzymes to detoxify contaminants.
- CO 6. Know about biological treatment of liquid wastes and solid wastes.

UNIT I (13 hours)

Environmental pollution: Types of pollution – Air, water, land, sound and radioactive pollution.

Water pollutants: Major sources- Domestic, municipal, industrial and agriculture; types and standards; Impact of water pollution on aquatic biota and human health.

UNIT II (13 hours)

Atmospheric Pollutants: Major sources, types and standards; Primary pollutants- Carbon monoxide, sulphur oxides, nitrogen oxides, particulate matter, hydrocarbons, asbestos and CFCs; Secondary pollutants; Impact of air pollutants on climate- Acid rain, photochemical smog, global warming, ozone depletion and haze.

UNIT III (13 hours)

Remediation: Types of remediation- Physical, chemical and biological; Bioremediation- *in-situ* and *ex-situ* bioremediation; Phytoremediation; Microbial remediation; Biological treatment of liquid wastes and solid wastes.

References:

1. Diwakar Rao, P.L. (1990). Pollution control Hand book, Utility Publications Ltd., Secunderabad, India.
2. Eaton, A.D., Clesceri L.S. & Greenberg, A.E. (1995). Standard Methods for the Examination of Water and Wastewater, APHA, Washington.
3. Moriarty, F. (1975). Pollutants and animals; A factual perspective. George Allan & Unwin Ltd., London.
4. Schmitz, R.J. (1996). Introduction to water pollution biology. Asian Books Pvt. Ltd., New Delhi.
5. Trivedi, P.R. and Sudarshan, K. (1995). Global Environmental issues, Commonwealth Publications, New Delhi.
6. Vernberger *et al.* (1981). Biological monitoring of marine pollutants, Academic Press, New York.
7. George, A. (2000). The Ecology of sea shores, CRC Press.
8. Agrawal, K.C. (2002). Environmental Pollution: Causes, Effects and Controls.
9. Binoda C. Sabata (1995). River Pollution in India.
10. Khetan S.K. (2000). Microbial Pest Control.
11. James, G.A. (1999). Ethical Perspective on Environmental issues in India.

BSE512 STEM CELL BIOLOGY AND REGENERATIVE MEDICINE

39hrs

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Gain in-depth knowledge in the field of stem cell biology and regenerative medicine
- CO 2. Understand the different types of stem cell and their applications.
- CO 3. Learn about state-of-the-art technologies, applications and ethics in research of stem cell biology.
- CO 4. Understand the legal and ethical aspects of stem cell research and applications.
- CO 5. Know the principles and applications of tissue engineering and nanotechnology.

Unit 1 (13 hours)

Basics of stem cell biology, origin, development, types and properties of stem cells, embryonic stem cells and induced pluripotent stem cells (iPSCs), foetal (amniotic, umbilical cord blood and stem cells from other embryonic tissues), adult stem cells- Hematopoietic stem cells, mesenchymal stem/stromal cells, neural stem cells, hepatic stem cells and skeletal muscle stem cells, cancer stem cells, state-of-the-art technologies, applications and ethics in research of stem cell biology and differentiation

Unit II (13 hours)

Introduction and principles of regenerative medicine, cell based therapies, pluripotency and regenerative medicine, Cell-cell interactions in tissue regeneration, Isolation and culture of stem cells, Viral and non viral vectors in stem cell research, Genome editing and use of genetically engineered stem cells. Applications of stem cell based therapies in bone, blood, cardiovascular regeneration, musculoskeletal repair, hepatocyte and neuronal transplantation, legal and ethical aspects of stem cell research and applications

Unit III (13 hours)

Nanotechnology: Definition, nanomaterials and their applications; Regenerative medicine, biomaterials and scaffolds in regenerative medicine, principles and applications of tissue engineering, modes of cell and tissue delivery, *in situ* tissue engineering and bioartificial organs, GMP and regenerative medicine

References:

1. Appasani K., Appasani, R.K.(2013) Stem cells and regenerative medicine, Humana Press, 2013
2. Lanza R., Atala, A. (2014) Essentials of stem cell biology, 3rd Ed., Academic Press

OPEN ELECTIVE COURSE
BSE 513 BEHAVIOURAL BIOLOGY

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Understand the evolution of social behaviour and types of social behaviours
- CO 2. Discern various types of social behaviours across organisms
- CO 3. Understand communication and the adaptive significance of these behaviours
- CO 4. Appreciate how epigenetics moulds behaviour

UNIT 1 (13 hours)

Sociobiology: Definition, introduction, history, scope, and significance. Basics of ecology and society; The evolution of animal societies. Instinctive, or intuitive behavior; Evolutionarily stable strategy hypothesis. Social behaviors: Aggregation, reproductive behaviour, territoriality, pack hunting, dominance interactions, parental care, and cooperative interactions within families.

UNIT II (13 hours)

Eusociality in insects: Hive society of social insects (ants, bees, and wasps); Eusociality in crustaceans (shrimps); mammals (mole rats); Cooperative breeding in birds, parental care. Social interactions in microbes - cooperation, conflict, and population. Spatial structure. Plant-pollinator networks.

UNIT III (13 hours)

Communication for social interactions: plumage, morphological characters, vocalizations, pheromones, vibrations; The adaptive significance of social organization; altruism; cooperation; courtship and reproductive behavior; the genetics, development, and epigenetics of social behavior.

References:

1. Aronson, E., & Aronson, J. (2018). The social animal. Worth Publishers, Macmillan Learning.
2. Brooks, D. (2012). The social animal: The hidden sources of love, character, and achievement. Random House Incorporated.
3. Buss, D. M. (Ed.). (2005). The handbook of evolutionary psychology. John Wiley & Sons.
4. Martin, P., Bateson, P. P. G., & Bateson, P. (1993). Measuring behaviour: an introductory guide. Cambridge University Press.
5. Peterson, G. R. (2005). Sociobiology: The new synthesis. 25th Anniversary edition.
6. Wilson, E. (2000). Sociobiology: the new synthesis, 25th anniversary edition.

III SEMESTER HARD CORE COURSES

BSH551BIOTECHNOLOGY

52hrs

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Understand how organisms and biological processes are used in biotechnology.
- CO 2. Gain in-depth knowledge of useful microorganism, bioreactors and fermentation technologies.
- CO 3. Learn about production of bioplastics, biofertilizers and biopesticides.
- CO 4. Learn about the application of plant tissue culture and production of various hybrid plants.
- CO 5. Become familiar with animal cell culture, production of transgenic animals and assisted reproductive techniques
- CO 6. Understand rDNA technology, principles and applications of PCR, RT-PCR and DNA fingerprinting.

UNIT I (13 hrs)

Aims and scope of biotechnology, basic concepts of biotechnology, traditional and modern biotechnology. Microbial Biotechnology: Fermentation techniques: Fermenters and bioreactors. Batch, submerged (SmF), solid substrate (SSF) and continuous fermentation. Fermented foods, oriental foods, silage, probiotics, single cell proteins, production of hormones and growth factors. Microbial polysaccharides, bioplastics, cell immobilization and its applications. Biopesticides (fungi, bacteria and viruses). Biofertilizers, plant-growth promoting microorganisms, biocontrol agents and bioprotectants. Transgenic microbes and their applications.

UNIT II (13hrs)

Plant Biotechnology: Plant tissue culture laboratory and aseptic techniques, culture media, callus induction, organogenesis, somatic embryogenesis, micropropagation, production of secondary metabolites, selective markers, somaclonal variation, synthetic seeds and cryopreservation. Haploid production: pollen, anther and ovule cultures. Cell suspension culture, protoplast culture, protoplast fusion and hybridoma technology. Transgenic plants, production of disease-, salinity-, pest-, herbicide-, drought-resistant and high yielding varieties of plants. Production of improved varieties using Ti plasmids. Application of rhizobia and mycorrhizas in plant tissue culture. Plant-derived vaccines and antibodies.

UNIT III (13 hrs)

Animal Biotechnology: Animal cell culture techniques, culture media, primary and secondary cell cultures, cell lines and cell strains and growth factors. Stem cells, gene expression in cell culture, organ culture, histotypic culture; Natural and synthetic cell culture media composition; cytotoxicity and cell viability assays; Transgenic animals and their uses. Animals as bioreactors. Assisted Reproductive Techniques: *In-vitro* fertilization, embryo transfer, super ovulation and cloning.

UNIT IV (13 hrs)

Molecular Biotechnology: Gene manipulation, restriction enzymes, DNA insertion through vectors, clone selection and expression of cloned genes. Expression systems and their applications: *Escherichia coli*, *Streptomyces*, yeast, baculovirus and animal cells as cloning

hosts. Analysis of DNA-DNA sequences, mutagenesis and gene expression, DNA extraction methods and amplification using PCR and RT PCR techniques; DNA fingerprinting. Overview of next generation sequencing and digital PCR highlights.

References:

1. Moo-Young, M. (2011) Comprehensive Biotechnology, Vol. 1, 2, 3 & 4, Pergamon Press
2. Cruger, W. & Cruger, A. (1990) A textbook of Industrial Biotechnology
3. Glazer, A. G. (1994) Microbial Biotechnology, WH Freeman and Co.
4. Peppler, H. J. (1979) Microbial Technology., Vol. 1 & 2, Academic Press
5. Bajaj, Y. P. S. (2007) Biotechnology in Agriculture and Forestry, Springer Verlag Publ.
6. Russell, G. E. (1988) Biotechnology of Higher Plants, Intercept Publ.
7. Reinert J., Yeoman, M. M. (1982) Plant Cell and Tissue Culture. A Lab manual. Narosa Publ.
8. Mantell, S. H. and Smith H. (1983) Plant Biotechnology. Cambridge Univ. Press.
9. Houdebine, L.-M. (2003) Animal Transgenesis and Cloning by John Wiley & Sons.
10. Butler, M. (2004) Animal Cell Culture and Technology, BIOS Scientific Publishers.
11. Davis, J. M. (2002) Basic Cell Culture: A Practical Approach (Practical Approach Series), Oxford university press, Oxford

SOFT CORE COURSES
BSS 552 ENVIRONMENTAL PHYSIOLOGY

39hrs

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Enhance the knowledge how the organisms are physiologically adapted to various environmental conditions.
- CO 2. Know the basic principles of plant responses to environment.
- CO 3. Understand the physiology of flowering, senescence and abscission.
- CO 4. Gain the knowledge about stress physiology; how the plants response to various biotic and abiotic stress. how plant adapted to the radiation environment.
- CO 5. Comprehend the physiology of circulation and respiration, including under special environmental conditions, such as high altitude and deep seadiving.
- CO 6. Know how some respiratory diseases are caused.

Unit I (13 hours)

Principles of plant responses to environment; Problems of environment; Ecotypes - the role of genetics. Photoperiodism and its significance, endogenous clock and its regulation and development. Physiology of flowering, Senescence- types, causes, physiology of senescence and its significance, Abscission.

Unit II (13 hours)

Stress physiology: Plant response to biotic and abiotic stress. Stress tolerance, heat resistance, HR and SAR, water deficit and drought resistance, salinity stress, metal toxicity, freezing and heat stress, oxidative stress; Plant adaptation to the radiation environment.

Unit III (13 hours)

Circulation: Types of heart and body fluids (blood and lymph); buffering properties of blood; blood circulation; Physiology and patterns of circulation; Circulatory physiological features in special environment viz., high altitude, deep seadiving. Respiration: Transport of oxygen and carbon dioxide; regulatory mechanisms of respiration, respiratory physiological features in special environments viz. high altitude, deep sea diving; respiratory diseases.

References:

1. Schmidt-Nielsen, K. (1981). *Animal Physiology Adaptations and Environment*. Cambridge University Press, Cambridge.
2. Prosser, C.L. & Brown (1983). *Comparative Animal Physiology*. W.B. Saunders.
3. Hoar, W.S. (1976). *General and Comparative Physiology*, 2nd Ed., Prentice Hall of India, New Delhi.
4. Wilson, J.A. (1979). *Principles of Animal Physiology*. MacMillan Pub., New York.
5. Hopkins, W.G. (1995). *Introduction to Plant Physiology*. John Wiley and Sons, Inc. New York.
6. Galston, A.W. (1989). *Life processes in plants*. Springer-Verlag, New York.
7. Nobel P.S. (1999). *Physico-chemical and Environmental plant physiology*, Academic Press, San Diego, U.S.A.
8. Taiz and Zeiser, E. (1998). *Plant physiology*. Wordsworth Publishing Co., California, U.S.A.
9. Baldwin, E. (1964). *An Introduction to comparative biochemistry* Cambridge Univ. Press, Cambridge.
10. Berne, R.M. & Levy, M.N. (1991). *Physiology*. The C.V. Mosby Company, St. Louis.
11. Ganong, W.F. (1971). *Review of Medical Physiology*, 5th Ed., Kotheri Book Depot, Bombay.

12. Guyton, A.C.& Hall, J.E. (1996). Text Book of Medical Physiology. 9th Ed. W.B. Saunders Company, Philadelphia.
13. Jenson, D. (1976). Principles of Physiology, Appleton Century Crafts.

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Gain in-depth knowledge in the field of developmental biology
- CO 2. Understand how gametes are produced, both in plants and animals.
- CO 3. Comprehend the process of cell differentiation at the molecular level.
- CO 4. Understand how the early developmental events occur in invertebrates.
- CO 5. Know how the genes play a role in axis specification and embryogenesis.

Unit I (13 hours)

Introduction: Chief events in animal development; History of thoughts and conceptual developments; experimental embryology; the concepts of differential gene activity. Gametogenesis in animals: Spermatogenesis; Oogenesis; Molecular events during fertilization. Gametogenesis in a few plant systems; early development in a typical plant.

Unit II (13 hours)

Cell differentiation: Definition and concept, Mechanism of gene action during cell differentiation; Factors influencing cellular differentiation. Creating multicellularity Cleavage types; gastrulation; Fate maps; Concepts of determination; Morphogenetic cell movements-cell adhesion and contact inhibition. Competence and induction, totipotency; Nuclear transfer experiments.

Unit III (13 hours)

Morphogenetic determinants in egg cytoplasm; Germ cell determinants and germ cell migration; Early vertebrate development-cell movements, Gastrulation, germ layers – ectoderm, endoderm and mesoderm. Neurulation and organogenesis; Developmental patterns in metazoans; Body axes - establishment of body axes in mammals; Genetics of axis specification in *Drosophila*; Homeobox concept - homeotic genes

References:

1. Davidson, E. H.(1976). Gene activity in Early Development. Academic Press. New York.
2. Browder, L.W., Erickson, C.A., Jeffery, W.R.(1991). Developmental Biology, 3rdEd. Saunders, Philadelphia.
3. Russo, V.E.A., Brody, S., Cove, D., Ottolenghi, S.(1992). Development - the Molecular Genetic Approach. Springer Verlag-Berlin.
4. Cartwright, T. (1994). Animal cells as Bio-reactors. Cambridge University Press, New York.
5. Malacinski, G. M. (1988) Development genetics of higher organisms, as primer in developmental biology. MacMillan Press, New York
6. Berrill, N.J. (1981) Developmental Biology. Tata McGrawHill.
7. Tyler, M. S. (2000) Developmental Biology: A guide for experimental study. Sinauer Associates, MA, USA.
8. Sussman M. (2011) Animal growth and development. PrenticeHall
9. Buttery P.J., Lindsay, D. B., Haynes, N, B.(1986) Control and Manipulation of animal growth.Elsevier, London.

PRACTICAL COURSES

BSP 554 BIOTECHNOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Develop laboratory skills in biotechnology
 - CO 2. Use solid surface fermentation technique for production of antibiotics.
 - CO 3. Carry out PCR and do the analysis
 - CO 4. Do vermicomposting and mushroom cultivation.
 - CO 5. Perform plant tissue culture techniques and check the nutritional and anti-nutritional qualities of edible seeds.
-
- 1. Production and analysis of vermicompost
 - 2. Identification, collection and cultivation of mushrooms
 - 3. Submerged and solid-substrate fermentation.
 - 4. Production and assessment of enzymes, mycotoxins, organic acids and antibiotics.
 - 5. Isolation and induction of root nodules by rhizobia
 - 6. Isolation and mass production of arbuscular mycorrhizal spores.
 - 7. Plant tissue culture
 - 8. Evaluation of nutritional and antinutritional qualities of edible seeds.
 - 9. Evaluation of soil qualities (e.g. texture, bulk density and water holding capacity)
 - 10. Evaluation of soil components (e.g. nitrogen, phosphorus, organic carbon)
 - 11. Pattern of decomposition of organic matter (e.g. leaf and woody litter)
 - 12. Biogas production
 - 13. Functional properties of food (e.g. water absorption capacity, gelation, foaming and emulsion)
 - 14. DNA extraction methods and PCR /RT PCR confirmation
 - 15. Analysis of RT PCR data in terms of copy number or quantification.
 - 16. Analysis of DNA and protein sequences.

BSP555 ENVIRONMENTAL PHYSIOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Conduct experiments in environmental physiology
 - CO 2. Determine blood indices, blood pressure and thermal stress.
 - CO 3. Demonstrate rate of transpiration, effect of temperature on the rate of respiration and plant responses to salinity and metal stress..
 - CO 4. Know how to check the seed health and effect of salinity on seed germination.
 - CO 5. Check viability of seeds, inducers and inhibitors of germination.
-
- 1. Haematology-
 - 1.1 Determination of blood indices
 - 1.2 Determination of blood pressure.
 - 2. Respiration-
 - 2.1 Estimation of oxygen consumption by the organism under stressed condition (thermal stress).
 - 2.2 Demonstration of rate of transpiration by photometry.
 - 2.3 Effect of temperature on the rate of respiration.
 - 3. Seed physiology–
 - 3.1 Seed health testing.

- 3.2 Determination of percent viability of seeds by germination method.
- 3.3 Germination inducers and inhibitors
- 3.4 Determination of β -amylase activity in germinating seeds.
- 3.5 Effect of salinity on seed germination.
4. Stress Physiology-
 - 4.1 Plant responses against salinity and metal stress
 - 4.2 Radioisotope methodology and its principles (GM Counter and Scintillation counter)

BSP 556 DEVELOPMENTAL BIOLOGY LAB

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Develop practical skills using model organisms in developmental biology
- CO 2. Gain the skills to isolate and mount the imaginal discs, sex comb, genital plate.
- CO 3. Carry out practicals on developmental mutants in *Drosophila* and *Arabidopsis*.
- CO 4. Carry out staining techniques for gametes and embryo.

1. Study of model organisms used in developmental Biology.
2. Isolation and mounting of imaginal discs.
3. Structure of sperms and eggs.
4. Isolation and mounting of sex comb and genital plate in *Drosophila*.
5. Study of developmental mutants in *Drosophila* and *Arabidopsis*.
6. Spiral cleavage and general development in snail.
7. Study of hemimetabolous and holometabolous development in insects.
8. Life cycle and metamorphosis in frogs.
9. Structure of *Drosophila* and chick egg.
10. Study of chick embryo by vital staining technique.
11. Developmental stages in frog.
12. Developmental stages in chick.
13. Study of spermatogenesis in rat.

BSP 557 PROJECT WORK

Course Outcomes:

After successful completion of the course, students will be able to:

- CO 1. Carry out a research-based study - select a problem, frame the objectives, conduct literature review, tabulate, represent and interpret the results.
- CO 2. Do field work for collection of samples, questionnaire-based surveys.
- CO 3. Apply research methodologies, techniques and tools to conduct lab- / field-based research
- CO 4. Understand different types of standard methods of citation and references.
- CO 5. Write the dissertation, present and interpret the research data scientifically.
- CO 6. Build up the capacity to carry out a research project independently.
- CO 7. Get skilled to be appointed/absorbed based on the theme of the project work.
