

MANGALORE UNIVERSITY
Department of Applied Zoology
MSc Zoology

ZOH453: ANIMAL CELL BIOTECHNOLOGY

Teaching Hours 10/Unit

COURSE OUTCOME

1. This course aims at inculcating the principles and skills of animal cell culture in students.
2. They are introduced basics of cell culture, cell lines primary cultures, types of cultures etc., media, maintenance of cultures types of cultures etc.
3. Different cell culture techniques including preparation of media, maintenance of cultures, sub-culturing, maintenance of records applications of cell culture are dealt in detail.
4. Culturing of embryonic and stem cells, Hybridoma technology and monoclonal antibodies their applications, commercial applications of cell culture are also discussed.
5. Course also introduces students to genetic engineering , plasmids, Cloning , methods of gene introduction to cells and animals, transgenic animals and genetically modified animals.
6. Students groomed in animal cell biotechnology course can be entrepreneur in cell culture based industry or can get into cell culture based research or can get employed in cell culture based industries .

UNIT I

Introduction, Historical perspective, advantages and limitations of animal tissue culture. Major differences in vitro .Types of tissue culture. Biology of cultured cells- Culture environment, cell adhesion, cell proliferation, differentiation, Initiation of culture, cell senescence, continuous cell lines. Design and layout of laboratory,-Construction, sterile handling area, incubation, culture room, service bench, Preparation, wash up, maintenance of sterile condition. Equipment- Essential, beneficial and useful equipments, consumable items. Culture medium- Physico-chemical properties, complete media, serum, serum free media, balanced salt solutions, selection of medium and serum. Preparation and sterilization- Apparatus, Reagents and media, storage, Contamination- Source and types of contamination

UNIT II

Primary culture- types of primary culture, Isolation of tissues – mouse and chick embryos, human biopsy material, Explant culture, primary cell culture, disaggregation- enzymatic, mechanical. Suspension culture. Cell lines: Definition, Evolution of cell lines, continuous cell lines, cell line designation, maintenance, subculture, maintenance records. Cell line banking, cryopreservation, cell viability assays. Lymphocyte culture technique and its applications

UNITIII

Culture and maintenance of human and mouse embryonic stem cells. Stem cells in gene therapies, stem cell based therapies for autoimmune diseases. Hybridoma technology – Cell hybrids, Production and application of monoclonal antibodies. Use of animal cells as replacement for whole animal in toxicity testing. Commercial application of animal tissue culture – Uses of animal cells in vaccine production

UNITIV

Genetic engineering-General introduction and concept, Transduction and transfection, C-DNA, Recombinant DNA techniques, Restriction enzymes, Salient features of cloning vectors, Different types of cloning vectors, Plasmids, Cosmids, Phagemids, Shuttle vectors, Viral vector. Outline of gene cloning, gene cloning procedures, C-DNA cloning, Gene libraries, Chromosome Walking and jumping, Recombinant selection and screening- genetic methods, immunochemical methods, South-western screening, nucleic acid hybridization, product recovery.

UNITV

Methods to introduce genes into animal cells, electroporation, viral vectors, retroviruses, lipofection, calcium phosphate co precipitation. Transferring genes into animal oocytes, Eggs and embryos – Transgenic animals. Use of transgenic technology in research, knock-out mice. Production of human disease equivalents in the mouse, Novel therapies for human diseases. Transgenic technology in the improvement of farm animals, transgenesis in animal cloning. Genetically modified (GM) plants and foods.

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